

1 **Setting up a methodology to distinguish between green oranges and leaves using**
2 **hyperspectral imaging**

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21

22 **ABSTRACT**

23

24 The estimation of green citrus fruit yield is a key parameter for growers and the industry.

25 The early estimation of orange yield at the immature green stage could influence the

26 future market price and allow producers to plan the harvest in advance, thus reducing

27 costs. This research can be considered as a preliminary step for designing low-cost

28 spectral cameras capable of being mounted on unmanned aerial vehicles (UAVs) to

29 estimate orange yield and defects. Images were acquired from oranges and leaves from

30 an orchard in Jeju island (Jeju, Republic of Korea), using two hyperspectral reflectance

31 imaging systems, one working in the range 400–1000 nm (visible/near infrared, Vis/NIR)

32 and the other between 900–2500 nm (short-wave infrared, SWIR). The main objective of

33 the research was to set up a methodology to select the relevant bands - from the two

34 spectral ranges studied - to distinguish between green oranges and leaves and to detect

35 defects, which will allow citrus yield to be estimated. Analysis of variance (ANOVA) and

36 principal component analysis (PCA) were used to select the key wavelengths for this

37 purpose; next, a band ratio coupled with a simple thresholding method was applied. This

38 study showed that the Vis/NIR hyperspectral imaging correctly classified 96.97% and

39 92.93% of the pixels, respectively, to distinguish between green oranges and leaves and

40 to detect defects, while with the SWIR system, the percentage of pixels correctly

41 classified for these two objectives were 74.79% and 89.31%, respectively. These results

42 confirm that it is possible to use a low number of wavelengths to estimate harvest yield

43 in oranges, which could pave the way for the future development of low-cost and low-

44 weight equipment for the detection of green and sound fruit.

45

46 **Keywords:** Orange; Harvest yield; Defect detection; Hyperspectral and multispectral

47 imaging

48 **1. Introduction**

49

50 The citrus sector is one of the most dynamic and important agricultural sectors.
51 This sector is of considerable economic value to the countries of the Mediterranean Basin,
52 China, Brazil, the United States and Southeast Asia (Republic of Korea), the main
53 producers of citrus fruit, in general, and of oranges in particular (FAO, 2017).

54 In the light of the economic importance of the orange on the international market,
55 it is of particular interest to obtain an estimate of the crop yield prior to harvesting, which
56 usually takes place when the fruits are of the same green colour as the leaves (Obenland
57 et al., 2009). In oranges, the fruits often reach physiological maturity and present excellent
58 eating quality while the peel is still green. Post-harvest de-greening practices are used to
59 speed up the fruit colour change and to make the fruit more acceptable for marketing,
60 since a shiny, yellow peel is what the market demands (Porat, 2008).

61 Therefore, tools are needed to identify these green fruits on-tree, to make possibly
62 decision on harvesting and to optimize the process, so that fruits of the highest quality are
63 picked in keeping with their subsequent industrial use.

64 Currently, the indexes used to determine quality in oranges are colour intensity
65 and uniformity, firmness, size, shape, quality of flavour, lack of decay and lack of defects
66 including physical damage (abrasions and bruising), skin blemishes and discoloration, as
67 well as insect damage (Arpaia and Kader, 1999): this last quality index is one of the most
68 influential factors on yield in fresh oranges (Leemans and Destain, 2004; Li et al., 2011).

69 These days, the large-scale measurement of these production and quality
70 indicators still constitutes a major difficulty for producers. On the one hand, it is believed
71 that drones with a sensing unit may be used in the near future to achieve this aim due to
72 the rise in their popularity for agricultural applications; on the other, hyperspectral
73 imaging (HSI) and multispectral imaging (MSI) are two emerging techniques which

74 could be used for this purpose, due to their ability to acquire both spectral and spatial
75 information and thus assess quality indexes in agricultural products (Dale et al., 2013).

76 As far as the present authors are aware, only two published studies have used HSI
77 technology to distinguish between green oranges and leaves on-tree. In the first, Kane and
78 Lee (2007) used an InGaAs camera with a spectral range of 900 to 1700 nm to identify
79 green oranges in the field. They applied a combination of three different spectral band
80 images to identify the green orange and achieved an accuracy of 84.5% in terms of
81 correctly classified pixels. In the other, Okamoto and Lee (2009) using a CCD
82 hyperspectral camera in the 369–1042 nm range applied stepwise forward variable
83 selection method and linear discriminant analysis was then developed with selected
84 variables - between 10 and 14 - to identify green citrus fruits in the field at different stages,
85 with detection accuracies for complete fruit ranging between 80 to 89%.

86 As regards detecting external defects in oranges, a number of studies have been
87 published to date, all of which were carried out under laboratory conditions, with the aim
88 of selecting the optimal wavelengths for this application with a multispectral system (Li
89 et al. 2011; Bulanon et al., 2013; Lorente et al., 2013; Li et al., 2016).

90 In addition, the recent development of small-sized hyperspectral and multispectral
91 sensors has enabled them to be attached to UAVs in order to obtain images with a high
92 spectral and spatial resolution (Dale et al., 2013). The use of multispectral instead of
93 hyperspectral cameras would reduce the cost of the system and would speed up the data
94 analysis (Kim et al., 2011), although it would be necessary to make a prior selection of
95 the optimal spectral bands for each particular objective.

96 Likewise, it is important to consider that the HSI measurement devices can affect
97 the quality of the image data, so the selection of the instrument and the wavebands play
98 an important role in optimising the performance of the application (Kim et al., 2011).

99 Most of the research carried out using multi and hyperspectral cameras to measure quality
100 attributes (sweetness and acidity, firmness, stage of maturity or detection of defects) in
101 fruit and vegetables have been carried out in the Vis/NIR region (400–1000 nm) (Kim et
102 al., 2011; Li et al., 2018), although other studies have also used instruments in the SWIR
103 range (1000–2500 nm) (Gowen et al., 2007). This spectral range is of particular interest
104 to the citrus sector, since it not only allows us to differentiate between specific areas of
105 the fruit (i.e. for detecting defects), but also includes the most suitable wavelengths for
106 measuring the chemical parameters, such as soluble solid content and acidity (Williams,
107 2001).

108 Thus, given the numerous options available in terms of equipment, the successful
109 implementation of hyperspectral or multispectral reflectance technology *in-situ* for crop
110 yield estimation requires the instrument to be selected previously and the correct
111 waveband combination to be found for this specific application. Although this aspect is
112 hugely relevant when designing low-cost and low-weight cameras, to our knowledge
113 there are no reports in the literature regarding the comparison between Vis/NIR and SWIR
114 HSI systems to identify green citrus fruits and to pick out defective ones.

115 The objective of this work was to evaluate – at the laboratory scale – two line-
116 scan hyperspectral reflectance imaging systems working in the Vis/NIR and SWIR
117 ranges, respectively, to estimate the crop yield of oranges based on the distinction
118 between green oranges and leaves and the detection of external defects (abrasions and
119 bruising, skin blemishes and discoloration) in the oranges. The performance of these two
120 systems was also compared and the optimal wavelengths were selected for the further
121 development of a low-cost and low-weight MSI system which could be mounted on
122 drones.

123

124 **2. Material and methods**

125

126 *2.1. Sampling*

127

128 The oranges and leaves were obtained from an orchard in Jeju island (Jeju,
129 Republic of Korea) in autumn 2017.

130 For the first of the objectives – to differentiate green oranges from leaves – 20
131 leaves and 24 green oranges were placed in plastic bags and immediately taken to the
132 laboratory. In the case of the leaves, wet tissues were put into the bags to maintain the
133 moisture content of the leaves. Once in the laboratory, images of the leaves and green
134 oranges were taken. Next, a set made up of 5 samples of green oranges together with their
135 leaves was measured to validate the model obtained.

136 For the second objective – to identify oranges with defects – a total of 20 oranges
137 with some external defects (abrasions and bruising, skin blemishes and discoloration)
138 were measured.

139 Although the number of samples available in this study could appear to be limited,
140 it is enough to evaluate the potential of the developed methodology.

141

142 *2.2. Hyperspectral imaging systems and image acquisition*

143

144 Two laboratory-based push-broom Vis/NIR and SWIR systems were used to
145 obtain the hyperspectral images of oranges and leaves. Details of both imaging systems
146 are given in Table 1.

147 The Vis/NIR system was made up of an Electron Multiplying Charge Coupled
148 Device (EMCCD) camera (Luca R DL-604M, 14-bit, Andor Technology, South Windsor,

149 CT, USA), a C-mount objective lens (F1.4 28-mm compact lens, Schneider Optics,
150 Hauppauge, NY, USA), a line scan imaging spectrograph (Vis/NIR, Headwall photonics,
151 Fitchburg, MA, USA) and halogen light sources (4 x 2 sets of 100 W) at a 45° angle to
152 the sample. Next, in order to reduce specular reflection, the system was equipped with a
153 glass rotating polarizer available for Vis/NIR region. The nominal reflectance range was
154 approximately 400 to 1000 nm, with a spectral resolution of 4.7 nm. In order to capture
155 the actual shape of samples by keeping the pixels shape nearly square, the number of lines
156 was set at 1200 and 1080, with the distance between the lines set at 0.250 and 0.278 mm
157 for the oranges and the leaves, respectively. Spectral data was stored on a 1200 x 502 x
158 128 hypercube for the oranges and a 1080 x 502 x 128 hypercube for the leaves.

159 For the SWIR system, a 384 x 256 pixel InGaAs camera (MCT, Headwall
160 Photonics, Fitchburg, MA, USA) with spectrograph (Headwall Photonics, Fitchburg MA,
161 USA) and C-mount 1.4/25 mm focal length lens (Navitar, Inc., Rochester, NY, USA) was
162 used to collect images over a wavelength range of 900 to 2500 nm with 6 nm spectral
163 resolution. The illumination for reflectance imaging was provided by six tungsten halogen
164 lamps (100 W) connected by optical fibres and set up at a 45° angle. Line-by-line images
165 were collected by a conveyor unit enable to cover the spatial shape of the samples; it was
166 set to move at a 0.328 mm/scan for oranges and a 0.364 mm/scan for leaves.

167 To obtain the imaging using each hyperspectral system, 3 oranges were placed on
168 a tray (30 x 12.5 cm) in a single row, while for the leaves, a batch of four samples on a
169 30 x 14 cm tray was imaged with a single take. The sound, green orange samples were
170 arranged so that the stems pointed upwards, whereas the defective oranges were manually
171 arranged to present the defects for imaging. As for the leaves, these were arranged with
172 the adaxial side facing upwards.

173 Tablet movement was controlled by the step interval and the number of steps.
174 Visual Basic 6.0 (Microsoft, Seattle, WA, USA) was used to run the HSI system and to
175 control both conveyor and motor speed (0.25 mm/s for Vis/NIR and 8 mm/s for SWIR),
176 instantaneous field of view (IFOV) and exposure time (Table 1). The two-dimensional
177 spectral and spatial data were captured by the EMCCD and InGaAs cameras and stored
178 in raw format as a 3D hypercube (two spatial and one spectral dimension), which
179 comprised each spatial location and spectral information at each wavelength (λ).

180 Due to the heterogeneous intensities of the light source across the whole
181 wavelength range, reflectance calibration was performed before each measurement by
182 taking dark and white reference images. The dark reference was obtained by covering the
183 camera lens with a black cap (0% reflectance) and the white reference by using white
184 Teflon (99% reflectance). The reflectance value (R) of the raw images (R_0) was calculated
185 using the dark (D) and white (W) reference and taking into account the correction factor
186 for the reference panel (C) as in following equation (Kim et al., 2001):

$$187 \quad R = \frac{R_0 - D_i}{W_i - D_i} \times C_i$$

188 where i is the pixel index ($i = 1, 2, 3, \dots, n$) and n is the total number of pixels and the
189 correction factor (C) of 1 was used.

190

191 *2.3. Image processing and analysis*

192

193 The steps followed in the full procedure of image processing and analysis for both
194 instruments are shown in Figure 1. To achieve this aim, Matlab software (version 2015a,
195 The Mathworks, Natick, MA, USA) was used. In the case of the SWIR system, the
196 spectral range used was 900 to 1900 nm due to the fact that there was no signal beyond
197 this wavelength.

198 After reflectance correction, the region of interests (ROIs) for sound green oranges
199 and leaves were manually selected using a reflectance image. The spectra of all pixels in
200 each ROI was extracted and averaged to obtain the mean intensity value for each
201 wavelength.

202 For the identification of oranges, the selection of the most significant wavelengths
203 for the distinction of green oranges and leaves was based on F-values of the analysis of
204 variance (ANOVA) between the two groups (i.e. green oranges *versus* leaves). The higher
205 the F-value, the more statistically significant the mean separation between groups (Neter
206 et al., 1996; Cho et al., 2013). In the case that only one wavelength could be extracted
207 from the ANOVA analysis, PCA was also used to determine the other wavelength needed
208 to obtain the ratio. Thus, the wavebands which presented the greatest difference were
209 used in the application of the ratio image.

210 For the detection of defects, PCA was used for all the hyperspectral data sets,
211 including the spatial and spectral dimensions. This algorithm reduces the spectral
212 dimensionality, since it converts the huge amount of data from the hypercube into a
213 limited set of scores and loadings. In this work, the PC images and the loading vectors
214 for the first three principal components (PC1, PC2, PC3) were used to select robust
215 wavelengths for the proposed objectives; for this goal, the mean centre was performed as
216 a pre-processing method (Wise et al., 2006).

217 Prior to using the PCA, a binary mask image was generated in order to avoid
218 interferences from background that could decrease the accuracy of the method. To
219 achieve this, the images at wavelengths 712.5 nm and 1065.11 nm were used for the
220 segmentation for the Vis/NIR and SWIR hyperspectral images, respectively, since they
221 showed the best contrast between the sample and the background. The background was

222 removed by setting a simple threshold value ($R < 0.045$ for Vis/NIR and $R < 0.073$ SWIR)
223 for these wavelengths, respectively, which was then applied to all the hypercubes.

224 Because the band-ratio can enhance the contrast between different regions (Vargas
225 et al., 2005), two different band-ratios were used to distinguish between the green oranges
226 and the leaves and to detect external defects in the oranges. The two-band ratio was
227 performed as the following equation:

$$228 \quad Q_{t/k} = \frac{R_t}{R_k}$$

229 where $Q_{t/k}$ represents a quotient of spectral reflectances, and R_t and R_k are the
230 reflectance intensities at t nm and k nm.

231 The frequency histograms of the ratio values were recorded in order to select the
232 optimal threshold values. Finally, the accuracy of the models was calculated as the
233 percentage of correctly classified pixels.

234 To ensure the robustness of the models developed, their external validation was
235 carried out. To achieve this, for differentiating between the leaves and the green oranges,
236 the 5 samples of oranges attached to leaves were used, while to detecting the defects, 25%
237 of the total defective samples (i.e. 5 fruits not used to develop the model) were randomly
238 selected for validation.

239

240 **3. Results and discussion**

241

242 *3.1. Spectral analysis*

243

244 Representative mean reflectance spectra of green oranges and leaves after
245 normalization, calculated from the pixel values of the ROIs for each system, are shown
246 in Figure 2.

247 Fig. 2a shows the mean spectra obtained using the Vis/NIR system in the spectral
248 region 400–1000 nm. Although the spectral patterns for both – green oranges and leaves–
249 were fairly similar, in the green-yellow area of the spectrum (500~600 nm) the average
250 reflectance obtained from the green oranges samples was higher than that obtained from
251 the leaves. This makes sense, since in the visible range, the dominant process taking place
252 is pigment absorption; in particular, the peak around 530 nm is due to β -carotene (Keşan
253 et al., 2016). In addition, one dominant spectral feature observed in both spectra is the
254 absorption of chlorophyll *a* at approximately 680 nm (Cho et al., 2013), whereas in the
255 range between 900 and 1000 nm, corresponding to the water band (Williams, 2001), the
256 oranges display lower intensity than the leaves.

257 The mean spectra obtained using the SWIR system in the spectral region 900–
258 1900 is shown in Fig. 2b. As in the case of the Vis/NIR system, the characteristic shape
259 of both spectra is very similar, with peaks and valleys in the same wavelengths. Here, the
260 greatest difference in reflectance values occurred mainly around 1100 nm and 1400-1600
261 nm. These values were related to the molecular vibrations corresponding to the C–H and
262 O–H bonds (Williams, 2001).

263

264 *3.2. Optimal wavelengths selection to distinguish between green oranges and leaves*

265

266 Figure 3 shows F-values for the ANOVA analysis between the two groups (i.e.
267 green oranges *versus* leaves) for each wavelength and the two systems tested.

268 With the Vis/NIR system, the F-values of each waveband in the range 400–1000
269 nm obtained from the ANOVA analysis for distinguishing oranges and leaves are
270 displayed in Fig. 3a. The highest F values were obtained for the bands 941.7 nm and 951.2
271 nm. Since these two bands were fairly close to each other, the band that yielded the highest

272 F-value was selected; in this case, the band 951.2 nm was chosen as one of the dominant
273 bands. This wavelength is very close to one of those chosen (967.2 nm) by Okamoto and
274 Lee (2009) to fulfil this objective using a sample group consisting of 3 varieties of green
275 oranges. This band corresponds to water absorption, which is the main component in
276 oranges (Williams, 2001).

277 Since only one wavelength could be extracted from the ANOVA analysis, PCA
278 had to be used to determine the other wavelength needed to obtain the ratio.

279 It was visually determined that the PC2 image (with 1.07% of the explained
280 variance) appeared to provide the best discrimination between green oranges and leaves.
281 The PC2 weighting coefficients showed high positive values in the red region and
282 negative values in the NIR region related to the O-H bond (Williams, 2001); the 698.2
283 nm wavelength, related to the absorption of chlorophyll *a*, was taken as the maximum,
284 dominant wavelength (Cho et al., 2013).

285 Based on these results, the 698.2 nm and 951.2 nm bands, obtained from the PCA
286 and ANOVA analyses, respectively, were selected for the MSI design. The raw and
287 binary images, as well as the image obtained after the application of band ratio $R_{\lambda 698}/$
288 $R_{\lambda 951}$ and its corresponding frequency histogram, are shown in Fig. 4.

289 To obtain the global classification capacity of the model, this band ratio was
290 applied to the validation set. The results indicated that for the $R_{\lambda 698}/ R_{\lambda 951}$ band ratio, the
291 highest classification accuracy (96.97%) for oranges *versus* leaves was obtained using a
292 threshold value of 1.00, which was obtained from the frequency histogram (Fig. 4d), in
293 which two clearly differentiated populations can be observed, with an overlap between
294 the intensity values of 0.90 and 1.00.

295 In the same way as in the Vis/NIR system, when using the SWIR system, the
296 dominant bands were selected from the F-values of ANOVA between the groups of green

297 oranges and leaves (Fig. 3b). The results showed that the wavelengths which gave rise to
298 a more significant separation were 1165, 1259 and 1471 nm.

299 Since the purpose of this study was to minimize the number of spectral bands so
300 that the measurement system to be developed would be as light weight, simple and
301 economical as possible, only two of these wavelengths were selected, those whose ratio
302 provided the greatest differentiation between leaves and fruits. Thus, from all the two-
303 band combinations available, the ratio of wavebands at 1165 and 1471 nm ($R_{\lambda 1165}/R_{\lambda 1471}$),
304 which corresponded to the molecular bonds C-H and the molecular vibrations caused by
305 O-H bonds, respectively (Williams, 2001), produced the clearest separation, as shown in
306 Fig. 5. These wavelengths coincide with two of those selected by Kane and Lee (2007)
307 for on-tree green citrus fruit identification, which is the only work with this objective
308 found in the literature.

309 The validation results showed that by applying a simple threshold value (2.60),
310 obtained from the frequency histogram (figure not shown), the classification accuracy
311 was about 74.79%.

312 The accuracy obtained using the SWIR system was around 22% lower than that
313 yielded with the Vis/NIR system. In Fig. 5, it can be seen that when applying the band-
314 ratio to the image with the SWIR equipment, not only is there a less distinct separation
315 between the leaf and the green orange than with the Vis/NIR system (Fig. 4c), but there
316 is also an area in the centre of the fruit with less intensity which corresponds to the
317 specular reflection caused by the geometry of the orange (Garrido-Novell et al., 2012).
318 With the Vis/NIR system, this difference was reduced by using the polarizer, whose
319 function is to reduce specular reflection in samples with curved or shiny surfaces;
320 however, this was not possible with the SWIR system, as this accessory is not easily

321 available for the SWIR spectral range. Thus, the problem is not the range of the camera
322 used, but the lack of a polarizer to reduce the effect of specular reflection on the fruit.

323

324 *3.3. Optimal wavelengths selection to detect external defects in oranges*

325

326 Fig. 6 shows the score images and loadings plots for the first three PCs obtained
327 in the calibration set of hyperspectral images of defective oranges after background
328 removal using the Vis/NIR system. The first three PCs accounted for 99.92% of the
329 explained variance, and it was found that the PCs above the third did not provide any
330 useful information for detecting defects (data not shown). In the image corresponding to
331 PC1 (96.37%), the areas of the oranges which especially stood out are those where, due
332 to the spherical shape of fruit, higher intensities were produced because they were closer
333 to the camera (Lee et al., 2008). Subsequent PC images represent other features ordered
334 according to variations in spectral responses. In general terms, PC2 images exhibit the
335 greatest contrast between the sound and defective areas of the oranges, and so appear to
336 have high discrimination power for identifying the defective areas. In PC3, the defects
337 can be observed, and the stem of the fruit can clearly be seen, although no visual
338 differences between these two features were recognized.

339 In addition, Fig. 6 also shows the loadings for the PC images (PC1–PC3) obtained
340 from the hyperspectral images across the Vis/NIR region. The peaks and valleys show
341 the dominant wavelengths, with maximums of 760 nm observed in PC1, 679 nm in PC2
342 and 665 nm in PC3, and minimums around 755 nm and 693 nm in PC2 and PC3,
343 respectively, with no minimum of note observed in PC1. In view of these results, it can
344 be stated that within the visible spectrum range, the red region and, in particular, those

345 wavelengths related to the absorption of chlorophyll a, are predominant (Martínez-
346 Valdivieso et al., 2014; Garrido et al., 2016).

347 Based on the visual aspect, PC2 is the component which seems to provide the best
348 detection of defective areas in oranges. Thus, based on the loading plot obtained for this
349 PC, the two most powerful spectral bands (679 and 755 nm) in PC2 were selected.

350 The resultant band ratio ($R_{\lambda 679} / R_{\lambda 755}$) was applied to the reflectance images, with
351 which the contrast between the sound surface and defects was more noticeable. After the
352 application of the mask, the threshold was established to isolate the defective surface.

353 However, given the high level of heterogeneity present in the samples, when the
354 validation of the model was carried out, the threshold value with which the highest
355 accuracy was reached was not the same for all the samples. As a result, to find an optimal
356 threshold value for separating sound from defective areas in oranges, the classification
357 accuracy was calculated with threshold values within the range 0.23–0.35 in an increment
358 of 0.02.

359 Fig. 7 shows the classification accuracy as a function of the threshold value
360 established. After analysing the results shown in Fig. 7, it can be concluded that the
361 highest accuracy (92.93% of the correctly classified pixels) was reached with a threshold
362 value of 0.35. These results were similar to those obtained by Li et al. (2011), who
363 selected bands 630 and 687 nm by analysing the principal components to detect 9 types
364 of defects in 'Navel' oranges, and, after applying the ratio, obtained a precision of 98.2%
365 in terms of correctly classified pixels.

366 To discriminate between defective and sound areas using the SWIR system, the
367 optimal wavebands were investigated using the same methodology used with the Vis/NIR
368 system. Thus, from the loading plot for PC2, the 1206 and 1518 nm wavebands were

369 selected, which are related with C–H y O–H absorptions, respectively (Williams, 2001).
370 The ratio image ($R_{\lambda 1206} / R_{\lambda 1518}$) was created, using 1206 nm and 1518 nm images.

371 According to Fig. 7, which shows the accuracy obtained for each threshold value,
372 the threshold value that yielded the best classification accuracy (89.31%) for the
373 validation set was 0.29.

374 For this second objective, the difference in the accuracy obtained by both systems
375 was not as great as in the differentiation between green leaves and oranges, and, the
376 Vis/NIR system enabled to obtain the model with the greatest accuracy.

377

378 **4. Conclusions**

379

380 The results obtained in this study indicate the feasibility of using HSI technology
381 to measure crop yield in oranges. The HSI systems can also potentially be developed
382 further as a low-cost multi-spectral imaging system using the key wavelengths identified
383 with the PCA method together with a simple ANOVA analysis from the calibration sets.
384 Four wavelengths (679, 698, 755 and 951 nm) could potentially be implemented as MSI
385 systems to differentiate green oranges from leaves and to detect orange peel defects,
386 respectively, in the Vis/NIR system, and four wavelengths (1165, 1206, 1471 and 1518
387 nm) could also be used for the same purpose using the SWIR device.

388 For the two objectives proposed in this study (identification of fruits and detection
389 of defects) in green orange, after using a two-band ratio coupled with a simple threshold
390 method, a comparison of the two hyperspectral devices produced a better classification
391 performance with the Vis/NIR system than with the SWIR system, with an accuracy of
392 96.97% when distinguishing between green oranges and leaves and an accuracy of
393 92.93% when detecting defects. Therefore, it could be concluded that Vis/NIR was the
394 most suitable system for this application, with the added advantage of the equipment

395 being more economical than the SWIR. However, it must be highlighted that the use of
396 the polarizer with Vis/NIR system improved the signal reducing the specular reflection
397 in samples, while for the SWIR system this accessory is not easily available. In addition,
398 it must be added that if, as well as estimating the crop yield, certain chemical quality
399 parameters in oranges also need to be measured simultaneously, it would be of great
400 interest to incorporate a band related to the absorption of water or glucides, which would
401 require the use of the spectral range of the SWIR system.

402 This work can be considered as a feasibility study and further studies are needed
403 for in field application of these systems. In this study samples were measured in
404 laboratory conditions using halogen lights and for remote sensing, under sun-light
405 illumination, other factors must be taken into account.

406

407 **Acknowledgements**

408

409 This work was carried out under the bilateral (Spain-Republic of Korea)
410 EUREKA R & D project entitled ‘Development of ICT fusion smart farm technology for
411 the intelligent production and distribution of oranges, INTELLIGENT-CITRUS’. The
412 authors wish to express their gratitude to the Spanish Ministry of Education, Culture and
413 Sports for the support offered to Irina Torres Rodríguez through the Training Programme
414 for Academic Staff (FPU).

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509
510

511 **Table 1**

512 Details of the two hyperspectral cameras used.

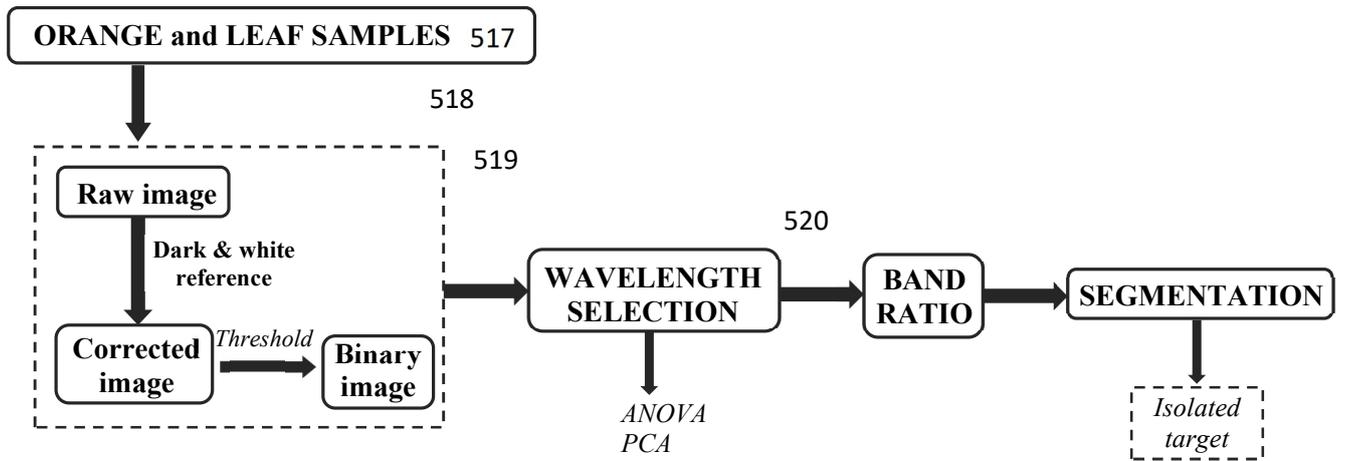
Wavelength range	Vis/NIR (400–1000 nm)	SWIR (900–2500 nm)
Manufacture	Andor Technology (South Windsor, CT, USA)	Headwall Photonics (Fitchburg MA, USA)
Sensor	EMCCD	InGaAs
Bit depth	14 bits	12 bits
Spatial resolution	8 μm	24 μm
Number of bands	128	275
Spectral resolution	~ 4.7 nm	~ 6 nm
Illumination	Eight 100 W tungsten halogen lamps	Six 100 W tungsten halogen lamps
Exposure time	10 ms	50 ms

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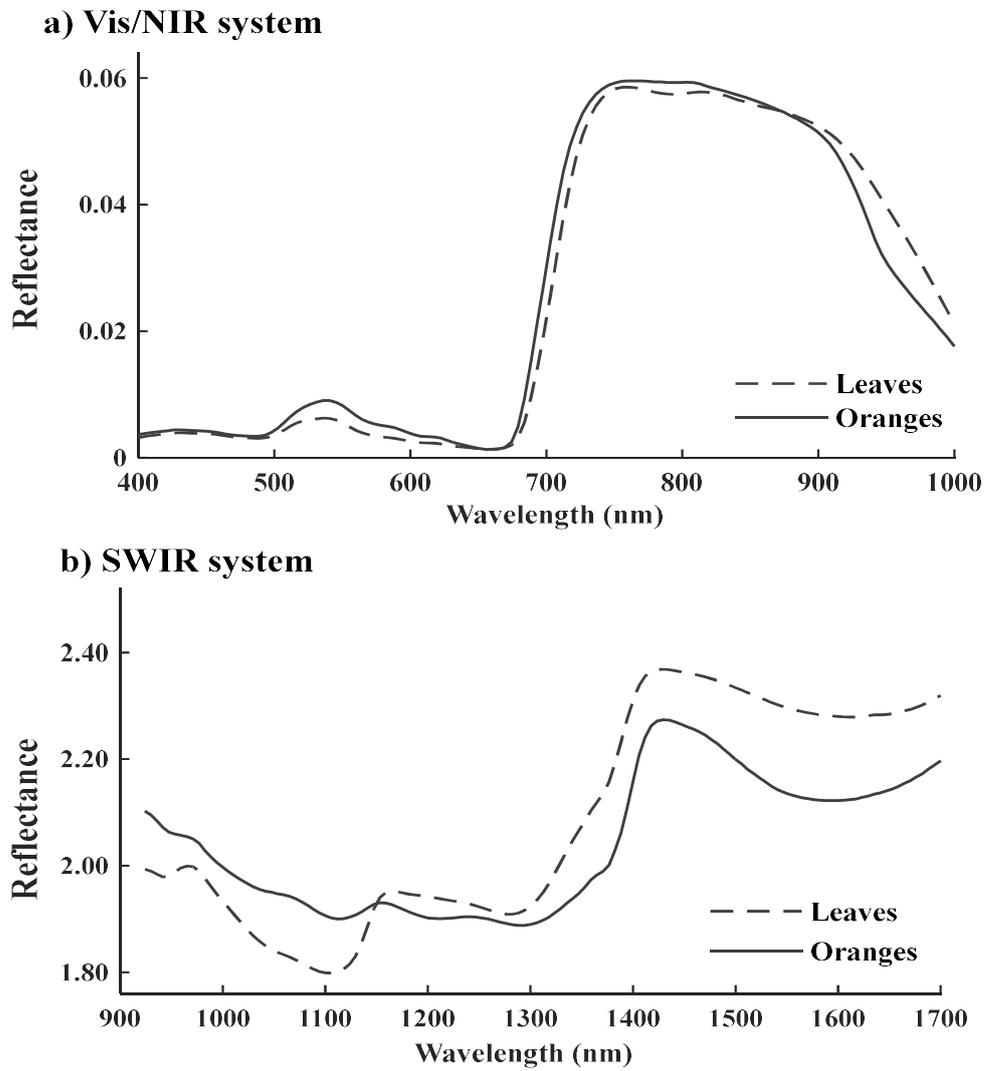
515 **Fig. 1.** Comprehensive flow for data analysis.

516



521

522 **Fig. 2.** Spectral features for leaves and green oranges obtained using the Vis/NIR (a)
523 and SWIR (b) hyperspectral imaging systems.

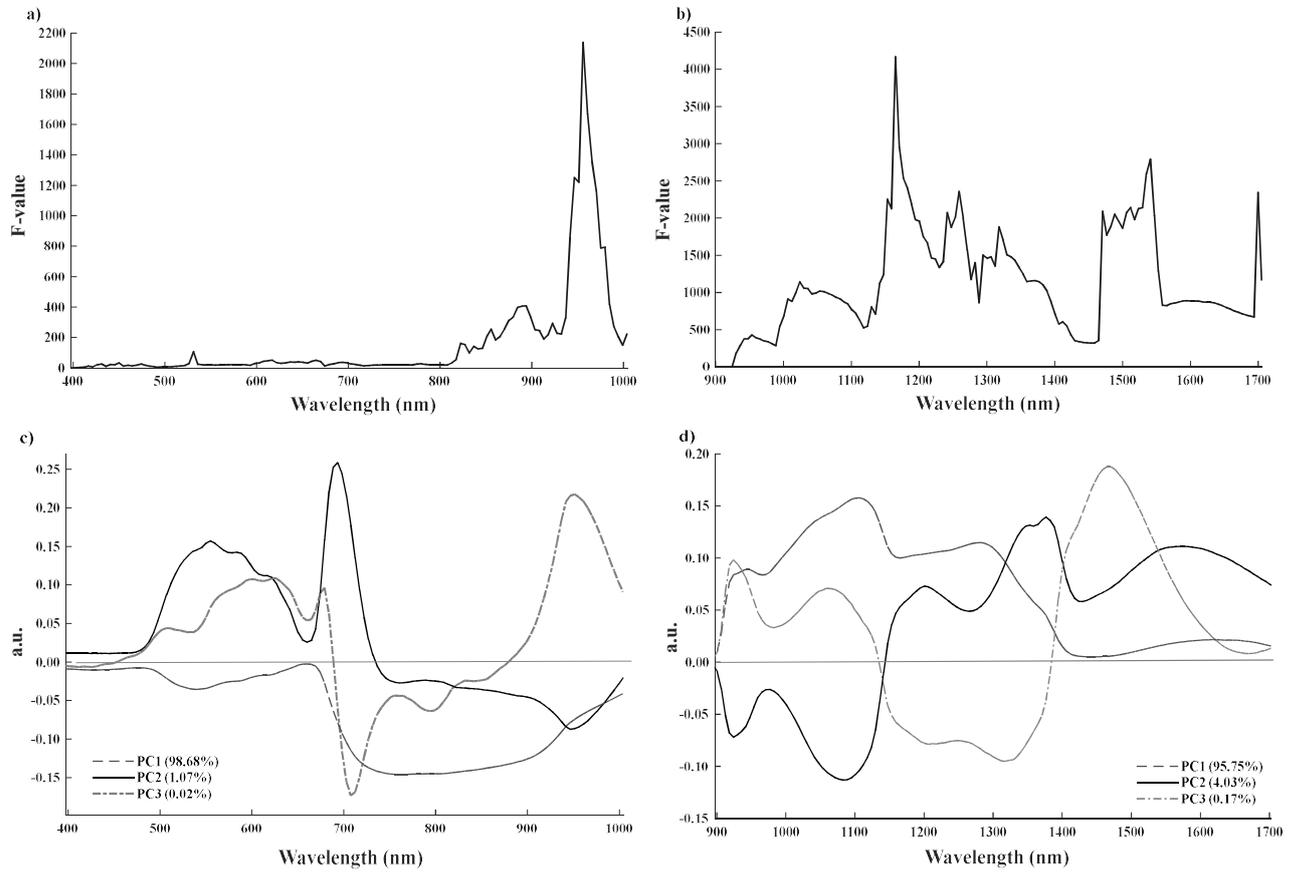


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527 **Fig. 3.** F-values obtained for the distinction between green oranges and leaves using the
528 Vis/NIR (a) and SWIR (b) hyperspectral imaging systems. Loading plots for the first three
529 principal components for the Vis/NIR (c) and SWIR (d) data sets.



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531

532 **Fig. 4.** (a) Reflectance image, (b) Binary image (mask), (c) Ratio image, (d) Histogram
533 of frequencies.

a) 534



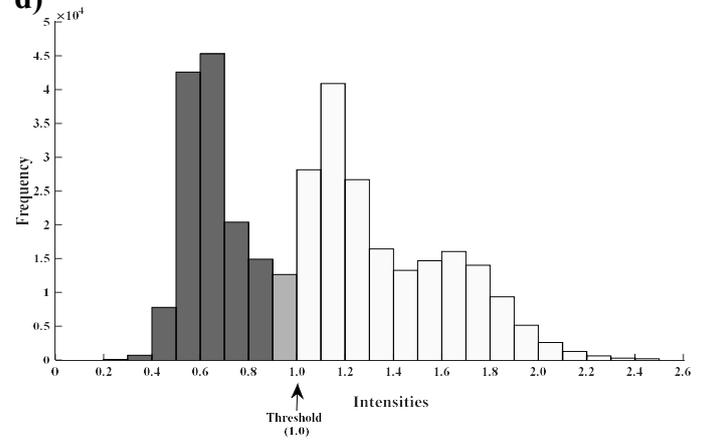
b)



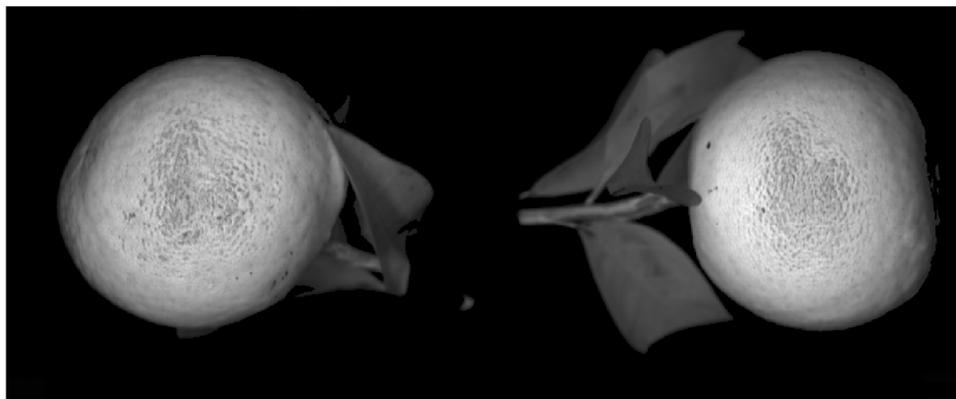
c)



d)



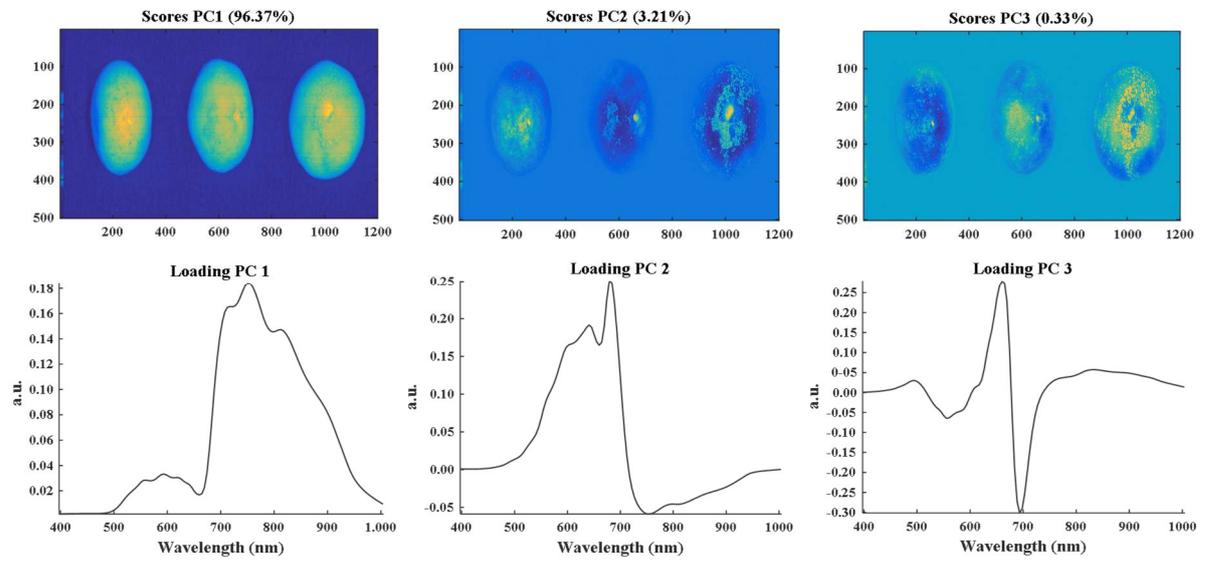
537 **Fig. 5.** Band ratio image ($R_{\lambda 1165} / R_{\lambda 1471}$) for the differentiation between green oranges and
538 leaves using the SWIR system.



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540

541 **Fig. 6.** PCA score images and loadings plot for the first three principal components
542 using the Vis/NIR system.

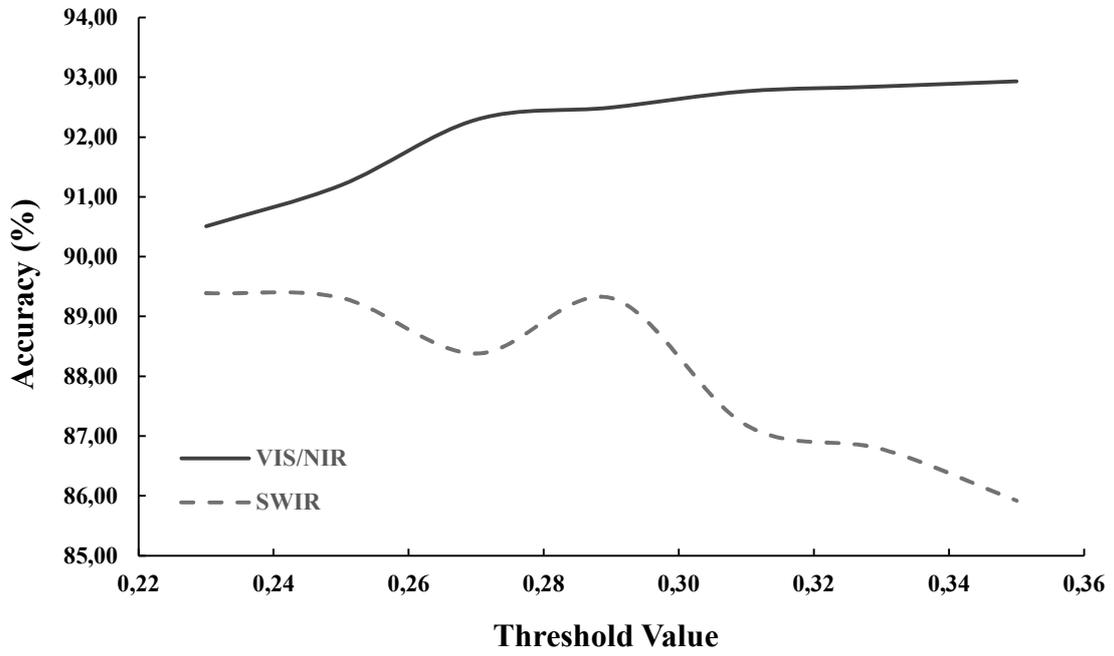


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545 **Fig. 7.** Accuracy (%) for each threshold value applied using the Vis/NIR and SWIR
546 based generated ratio images.

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