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## Capsaicin-Induced Thermal Hyperalgesia and Sensitization in the Human Trigeminal Nociceptive Pathway: An fMRI Study

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### Abstract

The aim of this study was to differentiate the processing of nociceptive information, matched for pain intensity, from capsaicin-induced hyperalgesic vs. control skin at multiple levels in the trigeminal nociceptive pathway. Using an event-related fMRI approach, 12 male subjects underwent three functional scans beginning 1 hour after topical application of capsaicin to a defined location on the maxillary skin, when pain from capsaicin application had completely subsided. Brush and two levels of painful heat (low - Thermal-1 and high - Thermal-2) were applied to the site of capsaicin application and to the mirror image region on the opposite side. Temperatures for each side were set to evoke perceptually-matched pain (mean temperatures [capsaicin/control]: Thermal-1=38.4/42.8°C; Thermal-2=44.9/47.8°C). We found differences in activation patterns following stimuli to treated and untreated sides in sensory circuits across all stimulus conditions. Across the trigeminal nociceptive pathway, Thermal-2 stimulation of hyperalgesic skin evoked greater activation in trigeminal ganglion and nucleus, thalamus, and somatosensory cortex than the control side. Thus, trigeminal nociceptive regions showed increased activation in the context of perceptually equal pain levels. Beyond these regions, contrast analyses of capsaicin vs. control skin stimulation indicated significant changes in bilateral dorsolateral prefrontal cortex and amygdala. The involvement of these emotion-related regions suggests that they may be highly sensitive to context, such as prior experience (application of capsaicin) and the specific pain mechanism (hyperalgesic vs. normal skin).

### Keywords

Pain; sensitization; dorsolateral prefrontal cortex; amygdala; S1; brush

Capsaicin-induced sensitization is considered to be a model for the hyperalgesic and allodynic states observed with neuropathic pain. Sensitization refers to the physiological phenomena of increased neural activity in response to a given stimulus intensity, while allodynia and hyperalgesia refer to perceptual symptoms. Applied topically or injected subcutaneously, capsaicin can produce a temporary state of behavioral hypersensitivity to thermal and

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mechanical stimuli in a healthy subject, similar to the chronic sensory symptoms of a patient with neuropathic pain (Baron, 2000; LaMotte et al., 1991; Simone et al., 1987; Treede et al., 1992). While capsaicin-induced sensitization has been demonstrated at the nociceptor level (Ren et al., 2005; Schmeltz et al., 2000; Serra et al., 2004), sensitization also occurs at spinal (Simone et al., 1991) and supraspinal levels (Lorenz et al., 2002). Numerous studies considering capsaicin-induced hyperalgesia and allodynia in the human have identified several brain areas with enhanced activity (Baron et al., 2000; Iadarola et al., 1998; Iannetti et al., 2005; Lorenz et al., 2002; Maihofner and Handwerker, 2005; Maliszka and Docherty, 2001; Witting et al., 2001; Zambreau et al., 2005).

However, these studies have not adequately addressed the influence of peripheral sensitization with respect to increased central activation with allodynic or hyperalgesic pain. Increased activation beyond first-order nociceptive neurons does not necessarily reflect central sensitization. A set stimulus level on hyperalgesic skin could produce increased afferent activity, and thus increased central activity as an intensity coding process. While previous studies have shown physiological changes supraspinally during hyperalgesia, the contribution of nociceptive circuitry preceding cortical processing in this state is unclear.

Previously, a positron emission tomography (PET) study attempted to use perceptually-matched stimuli to document changes in activation patterns in brain regions involved in the processing of thermal pain following capsaicin-induced allodynia (Lorenz et al., 2002). Relative to stimulation of normal skin, perceptually-matched thermal stimulation with capsaicin-induced heat allodynia revealed increased activity in medial thalamic nuclei, putamen, insula, anterior cingulate, prefrontal, and orbitofrontal cortices. These results indicated a dissociation between activity and perceived pain intensity, which they attributed to changes in cognitive and emotional responses to stimulation of capsaicin-sensitized skin. However, the supraspinal changes could have merely reflected increased afferent input with the presence of peripheral sensitization. The aim of the current study was to measure the relationship between peripheral and central activation when equally painful stimuli are presented to hyperalgesic vs. control skin.

The classical trigeminothalamocortical nociceptive pathway provides an opportunity to examine the effects of capsaicin-induced sensitization from the primary afferents to the cortex. It can be broken down into four main processing regions: the trigeminal ganglion (TG), subnucleus caudalis of the trigeminal brainstem complex (spV), thalamus, and the primary somatosensory cortex (S1). The trigeminal ganglion can encode noxious stimuli from the periphery, and sends this information to nociceptive neurons in the ipsilateral spV. The spV projects axons to the contralateral thalamus, where it relays nociceptive signals to S1 and other cortical structures. Somatotopic activation has been demonstrated in each of these regions with fMRI in response to noxious heat applied to the face (DaSilva et al., 2002; Borsook et al., 2003). The trigeminal pathway presents a practical model in which to measure sensitization along a nociceptive processing stream.

While the blood oxygen level dependent (BOLD) signal in the cortex has been correlated with slow varying field potentials (Logothetis et al., 2001) and action potentials (Thompson et al., 2003), the signal in the TG has not been defined. However, with stimulation, action potential firing presumably increases and the vascular innervation of the ganglion provides a substrate for BOLD signal changes. Potential issues related to measuring BOLD signals in TG have been discussed previously in a report describing somatotopic activations in the ganglion to heat and brush stimuli (Borsook et al., 2003).

The current study differs from the previous work in a number of important ways: (1) measurement of activity in trigeminal ganglion and the medullary dorsal horn using a region

of interest (ROI)-based approach, as well as a whole-brain analysis; (2) more closely matched pain intensities, using supra-threshold ratings rather than thresholds; (3) reduced scanner variability by contrasting capsaicin and control within the same scan; and (4) the increased spatial and temporal resolution of fMRI compared to PET. The null hypothesis was that there would be no differences in activation of nociceptive pathways following perceptually-matched painful heat applied to hyperalgesic vs. control skin. Any differences should reflect alterations in neural processing in categorization, transmission, and evaluation of peripheral inputs in non-sensory pathways.

## Methods

### Subjects

This study was approved by the McLean Hospital Institutional Review Board, and met the scientific and ethical guidelines for human pain research of the Helsinki Accord and the International Association for the Study of Pain. Seventeen healthy males were recruited through advertisements circulated in the Boston metropolitan area. All subjects provided informed consent to participate in this study. Three subjects were excluded following psychophysical screening (see below), one was excluded due to scanner problems, and one was unable to complete his fMRI session. The remaining 12 subjects averaged  $30 \pm 7$  years in age, and all but one were right-handed.

### Pre-scan Paradigm (Figure 1A)

**Capsaicin application**—Capsaicin cream (0.075% concentration; Zostrix, Rodlen Laboratories, Inc., Vernon Hills, IL) was applied topically to the left side of his face, within the receptive field of the maxillary division of the trigeminal nerve (V2). A dermatological bandage with a  $1.6 \times 1.6$  cm square cut out was used to limit the area directly exposed to the capsaicin cream, and a plastic film was placed over the application. A control cream (Lubriderm Sensitive Skin Therapy Moisturizing Lotion, Pfizer Consumer Healthcare, Morris Plains, NJ) was applied to the V2 area on the right side at the same time in a similar manner. The cream applications were left on for 25 minutes. Upon application of the creams, the subject alternately rated overall pain intensity and unpleasantness (see Supplemental Text 1) every 30 seconds using a computerized on-line visual analog scale (VAS).

The VAS was presented using the software package LabVIEW 5.1 (National Instruments Corp., Austin, TX) and the subject given a hand-held analog dial to rate his sensation. The scale was presented horizontally and labeled as “PAIN” or “UNPLS”, for pain intensity and unpleasantness, respectively. The markers on the extremes of the scale were “No Pain” and “Max Pain” for pain intensity, and “Min” and “Max” for unpleasantness. Subject ratings were stored digitally as numerical values from 0 to 10, though the VAS presented to the subject did not contain numbers. After the 25 minutes of cream application, the creams on both sides were thoroughly wiped off using a tissue, and the bandage was removed.

**Thermal stimulation**—All thermal stimuli were delivered using a  $1.6 \times 1.6$  cm contact thermode (TSA-II, medoc Advanced Medical Systems, Ramat Yishai, Israel). A homemade head cage, complete with two thermode holders, was created to fit around the MRI headcoil. One thermode was affixed to the capsaicin-treated (CAP) V2 area, and the other to the control-treated (CON) V2 area. At rest, the thermodes maintained a steady baseline temperature of  $32^\circ$  C. Stimuli were applied for 24 seconds, which included the rise time and return time to baseline. The effective rate of rise was  $5^\circ\text{C}/\text{sec}$ , while the rate of return was  $4.5^\circ\text{C}/\text{sec}$ . The inter-stimulus interval was 30 seconds.

After removal of the capsaicin and control creams using a tissue, an ascending series of temperatures was presented to each V2 area using 1°C increasing steps (Figure 1A). For each subject, two temperature levels reflecting minimal pain (Thermal-1: pain intensity VAS~2) and substantial, yet tolerable pain (Thermal-2: pain intensity VAS~7) were identified for CAP and CON V2. The temperatures selected for CON V2 were significantly greater than the temperatures for CAP V2 for both Thermal-1 [one-tailed paired t-test:  $t(11)=7.9329$ ,  $p<0.0001$ ] and Thermal-2 perceptual levels [ $t(11)=6.4590$ ,  $p<0.0001$ ]. The mean difference between the temperatures for CAP vs. CON V2 was  $4.4\pm1.5^{\circ}\text{C}$  for Thermal-1 and  $2.9\pm1.2^{\circ}\text{C}$  for Thermal-2. Subjects who showed no signs of thermal hyperalgesia (i.e. no difference in ratings for CAP vs. CON in response to the same temperature) were excluded from the study ( $n=2$ ). Additionally, one subject was excluded for being a poor discriminator of temperatures (could not discriminate between a  $40^{\circ}\text{C}$  and  $49^{\circ}\text{C}$  stimulus).

### Scanning Experimental Paradigm (Figure 1B)

**Stimuli**—Just over one hour after the capsaicin cream was removed, when pain due to the application had resolved completely, each subject underwent three fMRI scans, each focusing on one of three stimulus conditions, in this order: Thermal-1, Thermal-2, and Brush (Figure 1B). Each scan consisted of twelve 24 sec-on/30 sec-off stimulus cycles, with stimuli alternating between CAP and CON V2. The Brush stimulus consisted of 1Hz brushing of V2 along the superior-inferior axis. Brush stimuli were applied to the face using a lever system attached to a specially designed wooden frame overlying the MRI headcoil. Plastic/nylon materials allowed us to brush the face while standing at the feet of the subject.

A 24-sec rekindling thermal stimulus (the Thermal-2 stimulus for CAP V2, presented bilaterally) preceded the Thermal-1 scan, and was again presented prior to the Thermal-2 scan. This rekindling was performed to maintain capsaicin-induced hyperalgesia (Petersen and Rowbotham, 1999). A rekindling stimulus was not provided prior to the Brush scan, as the subject had been repeatedly exposed to the Thermal-2 stimulus before the Brush scan.

Every subject alternately rated pain intensity ( $n=3$  stimulus blocks rated/V2 side) or pain unpleasantness ( $n=3$  stimulus blocks rated/V2 side) using an online VAS scale projected onto a screen outside of the scanner. The subjects rated the first stimulus block with respect to pain intensity, and rated pain unpleasantness during the following stimulus block to the same V2 side. Subject ratings continued this alternating pattern until all 6 stimulus blocks were completed for each V2 side. As the act of rating strongly activates primary motor cortex contralateral to the hand being used, this could confound detection of activation in nearby S1 relating to stimulus application. Subjects were instructed to perform their rating with the hand opposite the side of stimulation, so as to separate dominant motor and somatosensory activations into separate hemispheres. The VAS scale also prompted subjects with which hand to rate each stimulus (LH or RH). In the event that the subjects felt no pain or unpleasantness during the brushing stimuli, subjects were told to move the VAS cursor to the center of the scale and to oscillate their rating with the brush strokes. This task was included to clarify that the pain vs. no pain contrast was not simply a rating vs. a lack of rating comparison. After the fMRI scans, subjects chose from a list of qualitative descriptors from the Gracely Pain Scale (Gracely et al., 1978) to report the sensory and affective intensity evoked by the stimuli.

### Data Acquisition and Analysis

**Image acquisition**—Imaging was realized using a 3T Siemens Trio scanner with a quadrature head coil. Anatomical images were acquired using a magnetization prepared rapid gradient echo (MPRAGE) sequence (128 1.33mm-thick slices with an in-plane resolution of 1 mm ( $256\times256$ )). Tissue-air interface areas can cause susceptibility effects, which lead to localized distortions in the EPI (functional) images. Such distortions can also arise from the

presence of the thermode on the face. Two series of images (T2 weighted Magnitude and Phase images) were acquired using a non-EPI spin echo sequence to estimate susceptibility distortions, and were used to correct EPI images utilizing FSL (FMRIB's Software Library, [www.fmrib.ox.ac.uk/fsl](http://www.fmrib.ox.ac.uk/fsl) (Smith et al., 2004)). For elaboration of the unwarping technique and theory, refer to its primary source (Jenkinson, 2001). Slice location, number, and thickness were the same as the ones used in the functional scans. Thermal-1, Thermal-2, and Brush functional scans were acquired as described in the paradigm (Figure 1B). Each functional scan consisted of 33 slices coronally oriented to match the brainstem axis and covering the middle region of the cerebrum. Slices were 3.5mm thick with in-plane resolution of 3.5 mm (64×64). A Gradient Echo (GE) echo planning imaging (EPI) sequence with TE/TR = 30/2000 was used to acquire the data. Three hundred and forty four brain volume images were captured for each functional scan, at the rate of one volume every 2 seconds. This resulted in a scan time of 11:28 per functional acquisition.

**Individual subject level image processing**—Functional imaging datasets were processed and analyzed using scripts within FSL. The initial four volumes were removed from each of the functional scans to allow for signal equilibration. Visual screening of the functional volumes revealed that none of the subjects showed indications of gross movement (>1 voxel). The skull and other non-brain areas were extracted from the anatomical and functional scans using FSL's script Brain Extraction Tool (BET). Motion Correction using FMRIB's Linear Image Registration Tool (MCFLIRT) was performed on each functional scan. The functional scans were unwarped using FMRIB's Utility for Geometrically Unwarping EPIs (FUGUE), and all volumes were mean-based intensity normalized by the same factor. The volumes were spatially smoothed with a 6 mm full-width at half-maximum filter, and a high-pass temporal filter was applied (sigma=54.0 sec). These functional images were then co-registered with the anatomical images using FMRIB's Linear Image Registration Tool (FLIRT).

First-level fMRI analysis of single subject data was performed using FMRI Expert Analysis Tool using FMRIB's Improved Linear Model (FEAT FILM) Version 5.4 with local autocorrelation correction (Woolrich et al., 2001). For the Thermal-1 and Thermal-2 scans, the temperature profiles recorded from the CAP and CON V2 thermodes were entered as separate explanatory variables (EVs), as were their temporal derivatives. The EVs for brushing stimuli were entered as boxcar functions. The EVs were convolved with a gamma function incorporating a 3 sec standard deviation and a 6 sec hemodynamic lag.

**Region of interest-based analysis**—A region of interest (ROI) based approach was used to trace activation changes along the classic trigeminal nociceptive pathway. A traditional whole-brain group analysis would dilute the signal in these regions, given the small size of many of these ROIs and inter-individual anatomical variation. This is particularly likely in the brainstem, where conventional transformations from original to standardized space consider primarily cortical references.

The trigeminal nociceptive pathway has been segregated into five main ROIs for the purposes of this study: TG, dorsolateral caudal brainstem (BS(c)), dorsolateral pontine brainstem (BS(p)), thalamus, and S1 (Supplemental Figure 1). These ROIs were chosen based on prior fMRI studies on the trigeminal nociceptive pathway (Borsook et al., 2003; DaSilva et al., 2002). The ganglia, brainstem, and S1 ROIs were individually hand-drawn for each subject based on their high-resolution anatomical scans (Supplemental Table 1). The ROIs for thalamus were identified in MNI space using *wfu\_pickatlas* (Lancaster et al., 2000; Maldjian et al., 2003), and transformed back into original space.

**a. Single Trial Averages (STAs)**—STAs were calculated using in-house programs implemented via MATLAB (Release 7.2, Mathworks Inc., Natick, MA, USA) in combination



with functional time courses and activation maps. For each subject, activation masks for each ROI were created based on uncorrected thresholding of the z-statistics ( $z > 1.64$ ) for both the CAP and CON condition for all three stimuli (Brush, Thermal-1, Thermal-2). The mean time course for each ROI was extracted from each subject's demeaned raw functional image, for each of the three stimulus types. Subjects without significant activation in a given ROI had their time course intensities set to zero. The EVs for each condition and stimulus were used to define each specific "trial". A trial was defined as the period consisting of 10 seconds prior to the beginning of the stimulus, and 50 seconds immediately following the beginning of the stimulus. This captured the duration of a stimulus response and its recovery without overlapping in time with the following stimulus. A trial average was calculated for each ROI, each stimulus, and each condition by taking the average time course of the trials. To measure a group STA response, the average and standard error at each time point of every subject's trial average was calculated.

**b. Logistic Regression**—For each stimulus type, logistic regression analysis was performed to assess the differentiability of two conditions, CAP and CON, based on activations (or contrasts of parameter estimates [COPEs]) within the selected ROIs (Pendse et al., 2006). A multivariate approach based on logistic regression attempts to distinguish the two groups based on all ROIs at once. This approach has several advantages: (1) it makes no assumptions about the distribution of the underlying data (such as normality); (2) likelihood ratio tests and Wald tests determine which variables are important for discrimination; and 3) an estimate of misclassification error is produced, which evaluates the model fit quality. ROI COPEs were sampled from significantly active voxels ( $z > 1.64$ , uncorrected), and zero values were substituted for subjects without significant voxels.

Logistic regression was used to determine the separability of CAP and CON, because nonparametric ROI-wise tests (e.g. Kruskal-Wallis) were not significant. The lack of a difference with a two-sample comparison would not suggest the statistical weakness of a multivariate approach, but rather a different statistical interpretation of the data. Since each ROI was not able to discriminate the two groups individually, this multivariate approach was used to determine whether the ROI set could discriminate these two states as a whole.

**Group level whole-brain image analysis (Figure 2A)**—Prior to group analysis, all first-level statistical maps were transformed to Montreal Neurological Institute (MNI) space. For each stimulus type, group activation maps were generated by fMRI expert analysis tool (FEAT) fMRIB's Local Analysis of Mixed Effects (FLAME) for CAP V2 and CON V2 separately. Statistical maps were thresholded using a mixture modeling approach to identify categories of activated and deactivated voxels (Everitt and Bullmore, 1999; Woolrich et al., 2005). In a conventional GLM, the same design matrix is used at every voxel in the brain with the noise as a multivariate Gaussian (Friston et al., 1995). However, incorrect modeling assumptions or unmodeled signals can result in a shifted and scaled null distribution. Recently, networks in the human brain have been identified that show similar patterns of activity even at rest (De Luca et al., 2005). Thus, under the null hypothesis, regions having similar patterns of activity can be expected to have differently scaled and shifted Gaussian distributions. This variability was accounted for by using Gaussian Mixture Modeling (GMM; Supplemental Text 2), a multiple comparisons-based analysis generally used for unsupervised classification of data into multiple categories (Pendse et al., 2006). A minimum cluster criterion of 7 voxels in original space (38 significant voxels in MNI space) was also implemented.

**Contrast analysis (Figure 2A)**—This analysis determined which regions were modulated after capsaicin application, relative to the side of stimulation. To directly compare mirror-like activations on our voxel-by-voxel approach, we flipped brains for control stimulation left to right in order to align them with the activations following stimulation to the capsaicin treated

side of the face. The use of flipped brains in fMRI analysis is a well-described procedure in clinical pain studies (Maihofner et al., 2006;Pleger et al., 2006;Schweinhart et al., 2006).

The parameter estimate maps for CON V2 stimulation were flipped along the y-axis to correspond with the stimulus lateralization of CAP V2 stimulation. That is, the statistical maps for the CAP and CON V2 condition were arranged such that areas contralateral and ipsilateral to stimulation corresponded in space. Paired contrast maps were separately generated for each of the functional scans. Significant differences were determined using the GMM method as described above.

To compare BOLD responses to stimulation of different sides of the face, a major potential confound needed to be addressed: hemispheric lateralized activity (Bingel et al., 2002, 2003;Brooks et al., 2002;Coghill et al., 2001). Hemispheric lateralized activity is a stimulus response that is evoked regardless of the side of stimulation (e.g. spatial attention). Such areas would confound a contrast using a brain flipping approach.

To reduce the likelihood of this possibility, a spatial mask was created to identify areas with hemispheric lateralized activity and to exclude these regions from the contrast analysis (Figure 2B; Supplemental Text 3).

## Results

### Capsaicin application

**Pre-scanning psychophysics**—The capsaicin application evoked painful and unpleasant sensations in all subjects (Figure 3A). Average maximum pain intensity reported was  $3.7 \pm 2.5$  (SD), with a range from 0.7 to 9.4. Average maximum unpleasantness was  $3.9 \pm 2.5$  (SD), with a range from 0.9 to 8.9.

**fMRI Psychophysics**—By the time of the beginning of the first scan, no subjects reported residual baseline pain from the capsaicin application. On-line perceptual ratings indicated that Thermal-1 and Thermal-2 stimuli (Figure 3B) were successful in evoking matched pain intensity levels in CAP vs. CON V2 during scanning (Figure 3C, Figure 4A). The reports during the Brush scan failed to show a significant difference between the two sides. The post-scan sensory and affective descriptors reported for each stimulus condition indicate that the qualitative sensations were comparable, though not identical, between CAP and CON V2 (Figure 4B, Figure 5).

### ROI-based analysis

**Single trial average responses**—STAs of the BOLD signal showed clear responses to each of the stimulus conditions across ROIs (example of responses in the S1 ROI, Figure 6A).

**Logistic regression**—Logistic regression multivariate analysis indicated that activation from stimulation of CAP and CON V2 could be distinguished on the basis of fMRI intensity values sampled across the five ROIs of the classical trigeminal nociceptive pathway (Supplemental Table 1). Z-statistic values (Wald's test) for all ROIs were significant, except for S1 with regards to Thermal-1 stimulation ( $z=0.3$ ). This indicates the participation of nearly every ROI in discrimination between group CAP and CON during brushing and thermal stimulation. Median changes of the ROIs in the two groups showed largely the same qualitative pattern for Thermal-1 and Thermal-2 (Figure 6B). Brush stimuli produced increased CAP responses in all trigeminal ROIs, with the exception of TG (Figure 6B).

## Whole-brain group fMRI results

With group-level analysis, thermal and brush stimulation to CON V2 produced activation and deactivation throughout the cortex, brainstem, and cerebellum. The painful thermal stimuli activated areas commonly identified in pain studies, including the insula (anterior, middle, and posterior), anterior cingulate cortex (ACC), S1, secondary somatosensory cortex (S2), and thalamus. Of note, deactivation was observed in the amygdala for Thermal-2, but not with the less painful Thermal-1 stimuli. Brush stimuli to CON V2 produced activation in areas found active in somatosensory studies, particularly insula and S2. Similar spatial patterns of activation were observed with stimulation to CAP V2 using Thermal-1, Thermal-2, and Brush.

**Contrast analysis**—When comparing activation patterns following perceptual intensity-matched stimuli, significant differences were found between responses to thermal stimuli delivered to CAP and CON V2 (Figure 7, Table 1). Thermal stimulation of CAP V2 elicited larger responses than CON V2 in contralateral frontal cortices (superior, middle, and inferior), ipsilateral cerebellum, and areas in or near ipsilateral amygdala (Figure 8). Decreased responses relative to control were found mostly opposite areas with increased responses, but also in the ipsilateral thalamus and inferior parietal lobe.

Brush stimulation of CAP V2 induced increased activation relative to CON V2 stimulation predominantly in contralateral frontal cortices (middle and inferior), contralateral posterior cingulate cortex, midcingulate cortex, bilateral amygdala, the paracentral lobule, ipsilateral cerebellum, and the brainstem (Figure 7, Figure 9). CAP V2 brush stimulation induced less activation than CON V2 stimulation in contralateral S1 (hand area) and primary motor cortex (hand area), and may be rating-related artifacts.

## Discussion

Capsaicin significantly alters responses to perceptually-matched thermal and brush stimuli in the trigeminal nociceptive pathway and in additional brain areas. ROI analysis revealed that BS(c) and Thal showed significantly greater responses with painful thermal stimulation of CAP V2 than perceptually matched stimulation of CON V2. Thermal stimulation of CAP vs. CON V2 also produced significant changes in TG and S1 that varied with perceptual intensity. In addition, brushing CAP V2 produced greater activation in BS(p), ACC, PCC, contralateral amygdala, and cerebellum than CON V2. All stimuli applied to CAP V2 produced significantly increased responses in the contralateral DLPFC (dorsolateral prefrontal cortex; Brodmann Area 9/46), the vicinity of ipsilateral amygdala, and ipsilateral cerebellum relative to stimulation of CON V2. Thermal stimulation of CAP V2 decreased responses relative to CON V2 in ipsilateral DLPFC, bilateral mid-insula, contralateral cerebellum, and areas adjacent to contralateral amygdala. This suggests that capsaicin application caused widespread changes in processing of sensory, affective, evaluative, and emotional dimensions of subsequent stimuli.

## Trigeminal Nociceptive Pathway

While some types of primary afferents can be sensitized by capsaicin (Ren et al., 2005; Schmelz et al., 2000; Serra et al., 2004), only Thermal-2 evoked increased responses in TG with stimulation of CAP V2 relative to CON V2, while Thermal-1 evoked a decreased response relative to control. The differential responses could be explained as a combination of peripheral sensitization and intensity coding. Since the capsaicin side produced less peripheral activation than control with Thermal-1, central sensitization could account for the similarity in perceived intensity. However, because lower temperatures were used for the capsaicin side relative to the control, a weaker response with capsaicin in the ganglia would not preclude the presence of peripheral sensitization.



Thermal and Brush stimulation of CAP V2 induced greater activity in BS(c). This area includes spV, which contains second-order neurons that receive nociceptive input from TG. Intradermal capsaicin sensitizes second-order neurons, increasing neuronal firing rates in response to thermal and mechanical stimuli (Dougherty and Willis, 1992; Simone et al., 1991). In this study, spV received enhanced input from the sensitized TG, and may further amplify these signals through second-order sensitization. Capsaicin-induced descending inhibition could also influence responses in spV (Hu and Sessle, 1979; Sessle et al., 1981), but was apparently insufficient to neutralize the increase.

All stimuli applied to CAP V2 significantly increased activation in thalamus, while Thermal-2 and Brush stimuli increased S1 activation. Previous imaging studies have detected increased stimulus-induced responses in S1 after capsaicin-induced thermal and mechanical allodynia (Lorenz et al., 2002; Maihofner and Handwerker, 2005; Maihofner et al., 2004). Single-unit neural ensemble recordings in rat indicate that capsaicin sensitization produces complex changes in the ventroposterior medial thalamus (VPM) and S1 (Katz et al., 1999). These changes were temporally distinguishable from alterations in spinal and brainstem nuclei, suggesting functional plasticity within VPM and S1. Such reorganization could increase BOLD responses in these areas.

While S1 activation has been suggested to encode pain intensity (Coghill et al., 1999; Moulton et al., 2005), CAP and CON V2 responses were significantly different with Thermal-2. Though Thermal-2 stimuli were matched perceptually, S1 responses were greater with stimulation of sensitized skin. However, Thermal-1 did not differentiate S1 activation with capsaicin and control conditions, and intensity-coding properties were observed with separate contrasts of Thermal-1 vs. Thermal-2 for CAP and CON V2 stimulation (unpublished observations). This suggests a dissociation between pain intensity coding and neural activity during hyperalgesia, when using more intensely painful thermal stimuli. Since the BOLD response represents a survey of overall neural activity within a voxel, including encoding of pain and touch (Davis, 2003), the differences in S1 during thermal hyperalgesia may represent a shift in the circuitry engaged (Katz et al., 1999). The pain intensity coding qualities observed previously are likely retained, but obscured by the recruitment of non-intensity coding neurons unmasked during thermal hyperalgesia. Alternatively, increased activation vs. matched perception could result from increased levels of activity in both excitatory and inhibitory neurons; balanced increases in these populations would result in no perceptual change, but would not necessarily be reflected in BOLD signal level.

An increase relative to control in BS(p) was identified for brushing of CAP V2. This area includes msn, which responds to afferents responsive to non-noxious mechanical stimulation. While msn has a large population of second-order low threshold mechano-sensitive neurons, it does not receive direct projections from small-diameter nociceptive afferents and contains few nociceptive responsive neurons (Kruger et al., 1988; Sugimoto et al., 1997). However, msn could receive modulatory input from spV (Greenwood and Sessle, 1976; Hallas and Jacquin, 1990), perhaps enhancing responses to mechanical stimuli with capsaicin exposure (Kwan et al., 1999). Alternatively, BS(p) contains other regions likely affected by capsaicin, including nucleus tractus solitarius and vagal nuclei.

### Beyond the Trigeminal Circuit

Areas of frontal cortex constituting DLPFC showed significantly altered responses to thermal and brush stimulation of CAP V2. DLPFC encodes cognitive control of complex actions (Fuster, 2001; Koechlin et al., 2003; MacDonald et al., 2000), and may affect pain perception through cortico-subcortical and cortico-cortical modulatory mechanisms (Apkarian et al., 2004; Lorenz et al., 2003; Maihofner and Handwerker, 2005; Wager et al., 2004; Zubieta et al., 2005). DLPFC is commonly active in pain imaging studies, and may participate in a proposed

pain-induced analgesia circuit (Anderson et al., 2002; Bencherif et al., 2002; Gear et al., 1999; Zubieta et al., 2001). This suggests that DLPFC changes during capsaicin-induced sensitization reflect an increase in its capacity to modulate nociceptive processes.

Capsaicin altered stimulus responses in or near the amygdala, including the uncus and portions of superior temporal gyrus (STG). This area processes the emotional salience of noxious stimuli (Becerra et al., 2001; Bernard et al., 1992), and is involved in pain-induced analgesia (Gear et al., 1999; Manning, 1998; Zubieta et al., 2001). Capsaicin-related changes in this study may indicate an enhancement of emotional salience of stimuli to CAP V2. This salience shift could initiate a tendency to protect an injured body part over an intact counterpart when faced with similar noxious input. Changes in amygdala responsiveness may reflect a change in the threshold for perceiving a stimulus as aversive.

The cerebellum was also more responsive to stimulation of CAP V2. Noxious stimuli may activate this area even in unconscious subjects, indicating an autonomic role (Hofbauer et al., 2004). The cerebellum may encode latent motor responses during pain (Peyron et al., 2000; Saab and Willis, 2003), and perhaps more importantly in the control of error signals (Ekerot et al., 1987). Thus, the differential responses here may relate to its potential role as a comparator for errors in somatosensory processing.

For thermal stimuli, increased DLPFC activation was observed contralateral to stimulation, and decreased activation on the ipsilateral side, and vice versa for the amygdala and cerebellum. Two possible explanations for this observation are: (1) these areas show functional lateralization with respect to stimulus side, which is altered with thermal hyperalgesia, or (2) hemispheric lateralized processes were inappropriately flipped in the contrast analysis. In addition to analysis steps that minimized the possibility of the latter, the contrast results for either side were spatially asymmetric. Also, statistical maps for both CAP and CON V2 stimulation produced bilateral activation in these areas. This suggests that the bilaterally opposite contrast results represent real changes in stimulus-based lateralized activation.

Mechanical allodynia was not detected in this study. This is perhaps explained by the long period of time (>1.5 hours) between the removal of the capsaicin and the brush functional scan. Despite the use of a rekindling stimulus (Petersen and Rowbotham, 1999), mechanical allodynia may have faded to non-painful levels during this time. Though dynamic mechanical allodynia has been reported to last up to 4 hours using a heat/capsaicin sensitization model (Dirks et al., 2003; Petersen and Rowbotham, 1999), the capsaicin model used in the current study may not have been sufficient to maintain allodynia over this time.

Despite the lack of psychophysical evidence for mechanical allodynia, brushing capsaicin-treated skin evoked increased activation in the same areas as heat, and also in ACC, which encodes pain intensity and/or unpleasantness (Buchel et al., 2002; Coghill et al., 1999; Derbyshire et al., 1997; Porro et al., 1998; Rainville et al., 1997), and PCC, which activates in chronic pain patients (Derbyshire et al., 2002; Hsieh et al., 1995; Lorenz et al., 1998). Even in the absence of perceptual changes, this data suggests that the brain changes after capsaicin exposure. These physiological changes may reflect a pre-symptomatic phase in the early development of progressive tactile allodynia (Ma and Woolf, 1996). Alternatively, as this study was not controlled for order effects, the activation patterns observed may be affected by this factor. The contralateral-ipsilateral contrast patterns in the brushing scan may thus be a result of the order of stimulus presentation.

### Perception Dissociated from Neural Activity

Differences in brain activation were found with perceptually-matched stimuli under conditions of thermal hyperalgesia. Although one study has acknowledged the presence of this

dissociation between brain activity and perception with thermal allodynia (Lorenz et al., 2002), its implications have not been discussed. This dissociation can be interpreted several ways. First, the perceptions may not have truly matched (Casey, 1996). Though VAS ratings were matched, there were nonetheless subtle qualitative differences in the perceptual experiences (Figure 5). Qualitative aspects of sensation are differentially coded throughout the brain (Strigo et al., 2003), and have been demonstrated with microstimulation within ventral caudal thalamus (Patel et al., 2006). Slight qualitative differences in sensory experience may be responsible for the differential responses following capsaicin-induced sensitization. Second, variability in subjective pain experiences has been attributed to supraspinally-mediated factors, such as expectation, psychological factors, and experience (Coghill et al., 2003). Perhaps during perceptually-matched stimulation, such cognitive factors occur but without significantly altering sensory processing. Third, evidence suggests that nociceptive inputs entering through separate channels (peptidergic and nonpeptidergic nociceptors) may be differentially affected by capsaicin (Braz et al., 2005). Nonpeptidergic neurons ascend to regions such as the amygdala, and may explain the observed differences. Furthermore, activation in both classes may be additive, accounting for differences in afferent systems at the level of the TG and BS(c).

## Conclusions

This study demonstrates that capsaicin alters nociceptive processing at multiple levels along the trigeminothalamocortical nociceptive pathway, from ganglia to cortex. Such changes may reflect a recalibration of descending inhibition and ascending nociceptive processes, in addition to plasticity in emotional processing. With Thermal-1 in particular, matched pain levels coupled with decreased ganglia activation with capsaicin relative to control indicate the presence of central sensitization. Further investigation of these kinds of changes should contribute to our understanding of clinical pain conditions with thermal hyperalgesia, such as migraine or neuropathic pain.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgements

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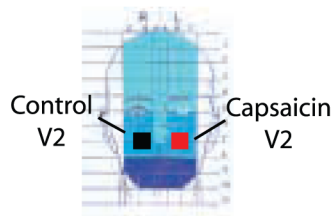
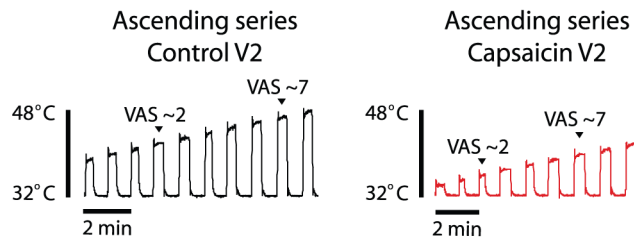
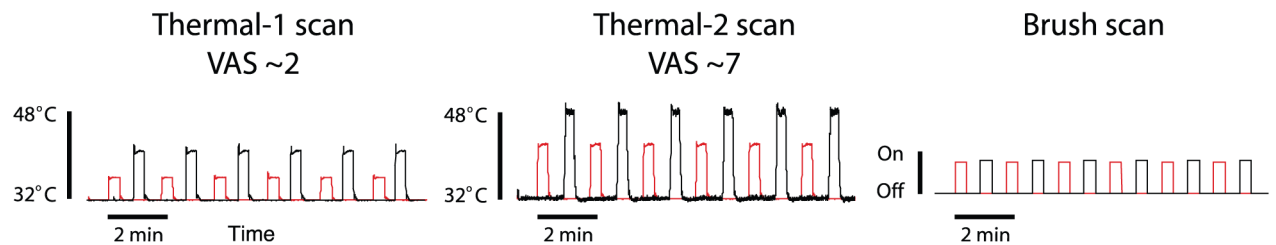
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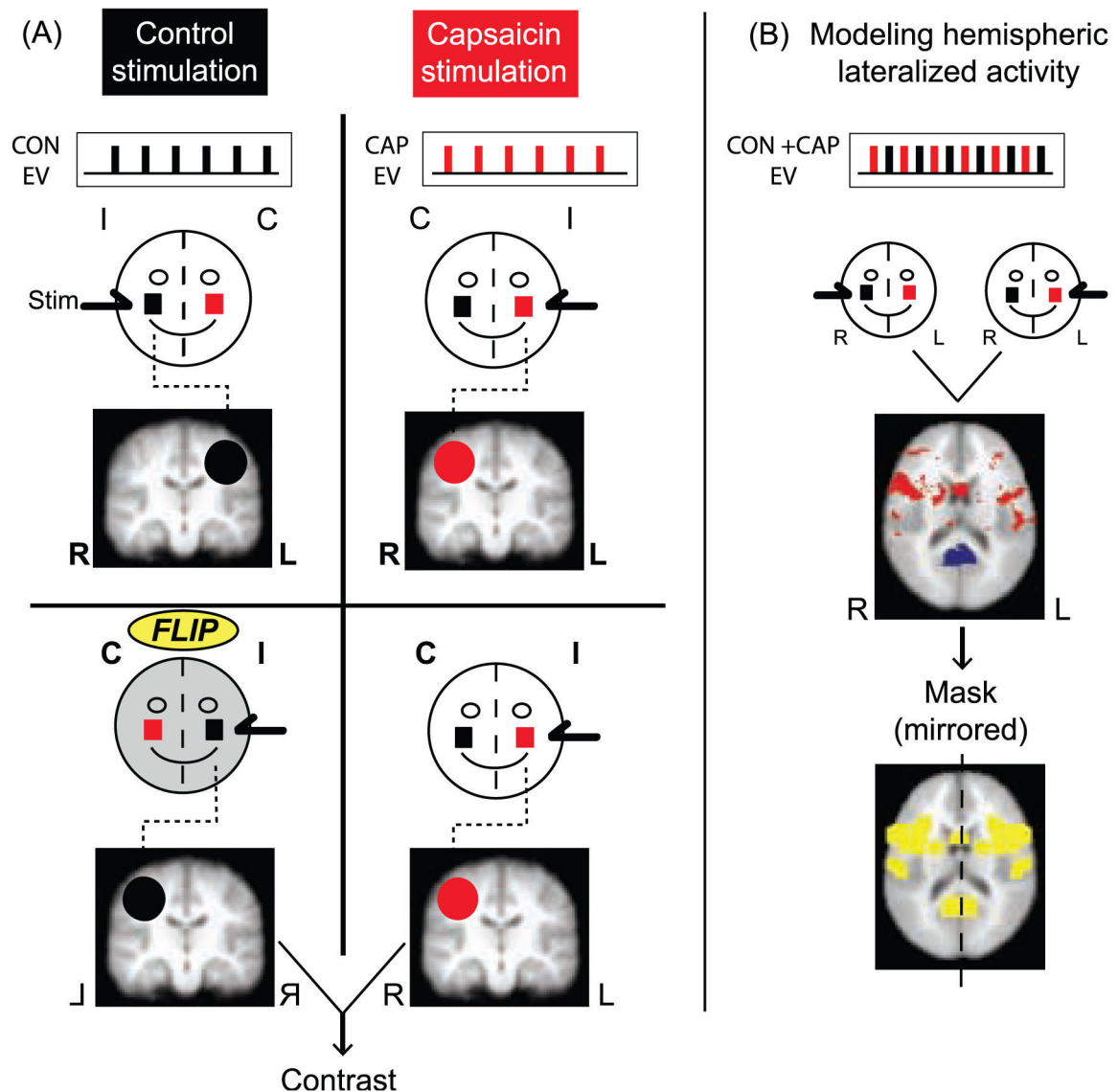


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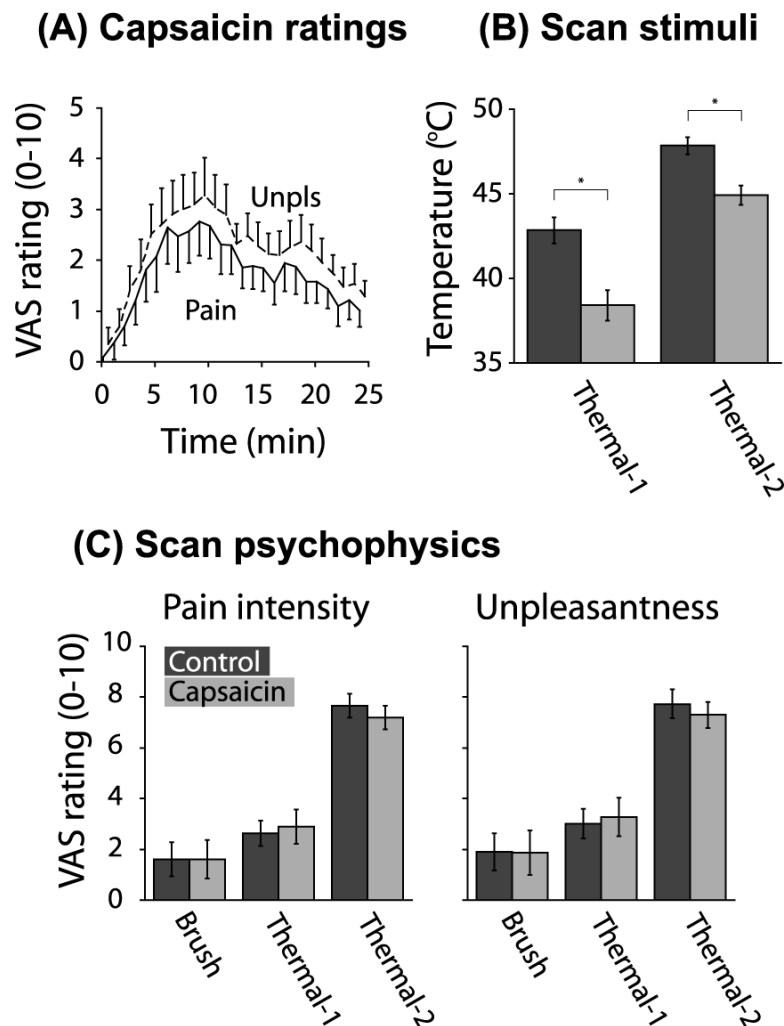
**(A) Pre-scan paradigm****Capsaicin application  
25 min****Subject-specific temperature calibration****(B) fMRI scanning****Figure 1.**

**Experimental paradigm.** A) Pre-scan testing. For 25 minutes 0.075% capsaicin cream was applied to the left V2 area (CAP V2), and a control applied to the right side (CON V2). For each side of the face, an ascending series of temperatures was presented to customize stimulus levels of Thermal-1 (pain intensity VAS= $\sim$ 2) and Thermal-2 (VAS= $\sim$ 7) for each subject. After the ascending series on CAP V2, a single Thermal-2 stimulus was applied to CON V2, to verify that the temperature for Thermal-2 on CON V2 and the temperature for Thermal-2 on CAP V2 felt similar to the subject. B) Stimulus delivery during fMRI scans. Each graph depicts stimulus delivery alternating between CAP V2 (red trace) and CON V2 (black trace). The Thermal-1 and Thermal-2 scans consisted of within-scan perceptually-matched temperatures, while the Brush scan used physically identical brushing stimuli. The scans were performed in the order presented here.



**Figure 2.**

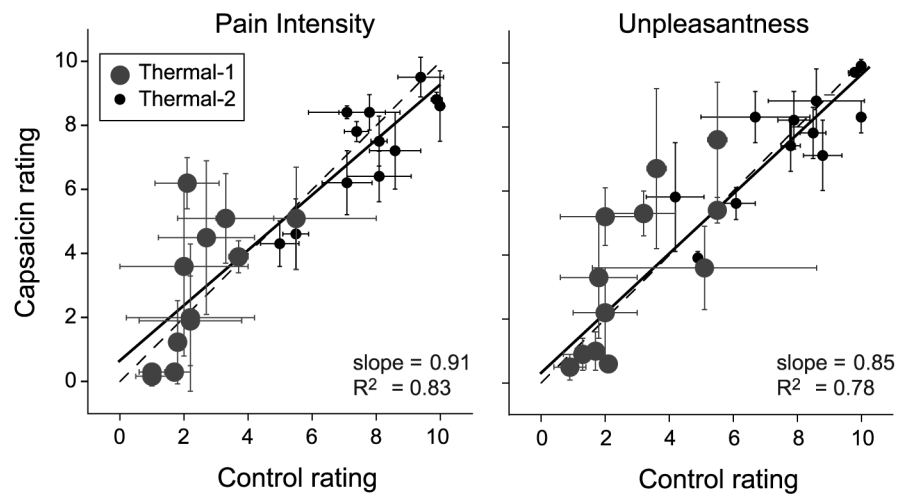
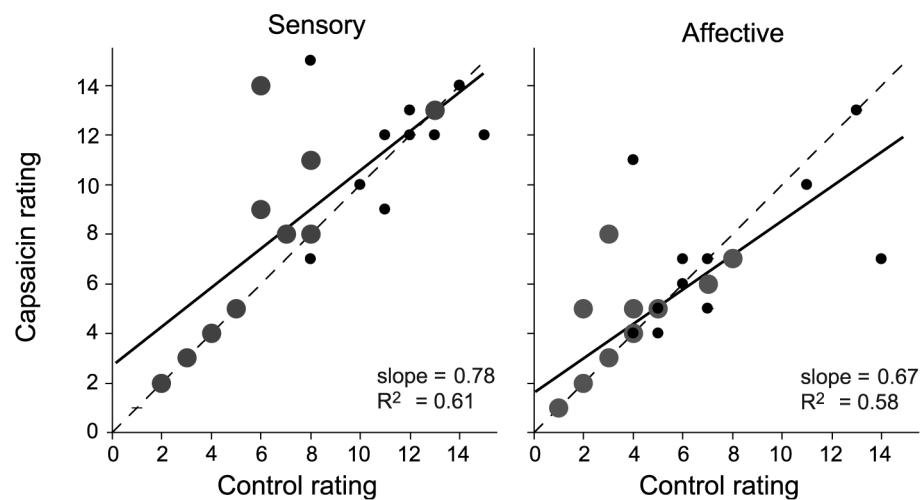
**Whole brain analysis approach.** A) Hemispheric flipping procedure for the contrast of CAP vs. CON V2 activation. Circles represent the subject's face, and the red square indicates the capsaicin site. I: ipsilateral to stimuli, C: contralateral to stimuli, L: left, and R: right. Beneath each stylized face are the explanatory variables (EVs) for the stimulus applied, and a schematic of its corresponding activation map. The second row of figures show the control activation map being hemispheric flipped along the y-axis, so that the brain areas contralateral to stimulus application are coincident. The contrast is performed between these reoriented images. B) Modeling hemispheric lateralized activity. A general linear model analysis creates an activation map based on stimulus application (regardless of the side of the face being stimulated). The original orientation for these images is maintained for both (i.e. not flipped). Red indicates activation and blue is deactivation. The second brain image shows the conversion of hemispheric lateralized activity into a spatial mask (in yellow). This mask reflects a mirroring of the hemispheric lateralized activity from the image above it. This mask is applied to the image from the contrast analysis, thereby creating a conservative contrast image. This is meant to correct for possible artifacts created by the flipping process.



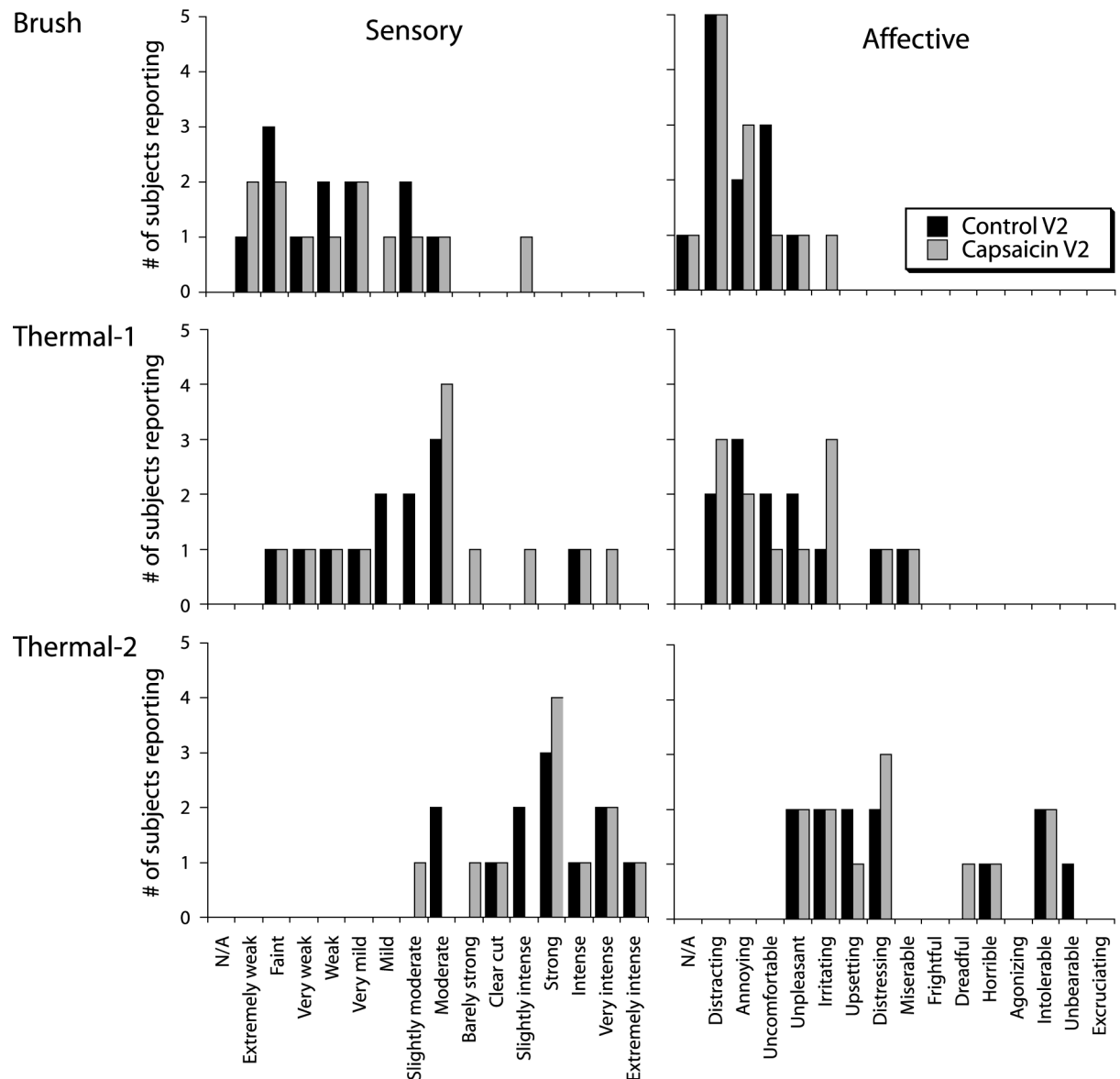
**Figure 3.**

**Psychophysical and stimulus data.** A) Group on-line ratings of pain intensity and pain unpleasantness during capsaicin application. The cream was applied at time 0. Mean $\pm$ SE. B) Mean temperatures applied to CON vs. CAP V2 for Thermal-1 and Thermal-2. Mean $\pm$ SE. \*One-tailed paired t-test,  $p < 0.0001$ . C) Group psychophysical ratings obtained during the Brush, Thermal-1, and Thermal-2 scans. The Brush psychophysical data from one subject was lost due to a computer error. Pain intensity ratings – the maximum pain intensity perceived was recorded per stimulus event, averaged across like events, and averaged across subjects. No significant difference in pain intensity was observed between CAP and CON V2 during the Brush [two-tailed paired t-test:  $t(10)=0.0765$ ,  $p > 0.1$ ], Thermal-1 [ $t(11)=0.9031$ ,  $p > 0.1$ ], and Thermal-2 scans [ $t(11)=1.9751$ ,  $p > 0.05$ ]. Unpleasantness ratings – the maximum unpleasantness perceived was recorded per stimulus event, averaged across like events, and averaged across subjects. No significant difference in unpleasantness was observed between CAP and CON V2 (two-tailed paired t-test) during Brush [ $t(10)=0.3606$ ,  $p > 0.1$ ], Thermal-1 [ $t(11)=1.2562$ ,  $p > 0.1$ ], and Thermal-2 scans [ $t(11)=0.6717$ ,  $p > 0.1$ ]. Mean $\pm$ SE.

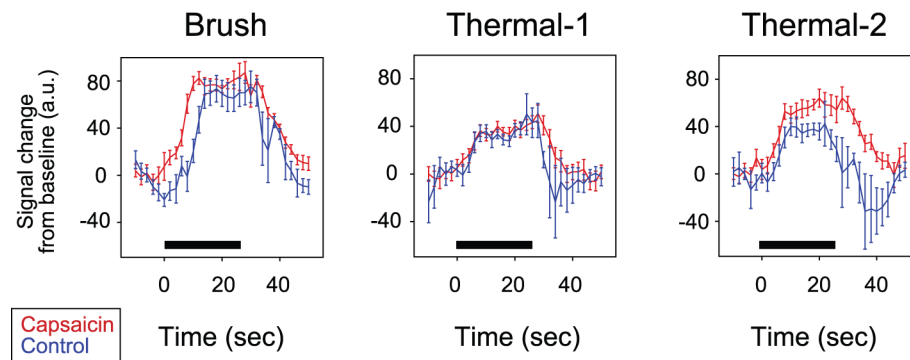
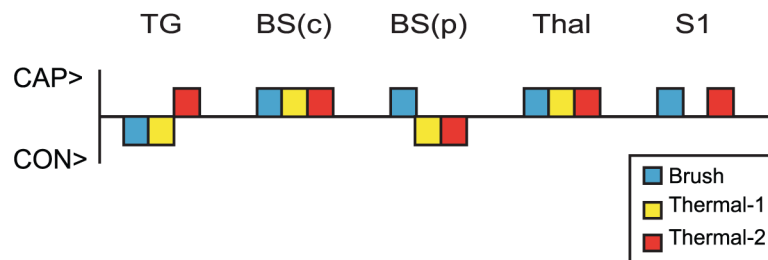


**(A) Online VAS Ratings****(B) Post-scan Gracely Ratings****Figure 4.**

**Within-subjects comparison of capsaicin vs. control ratings.** A) Online VAS ratings (0-10) for pain intensity and unpleasantness during scanning. Each dot represents an individual's response to a stimulus applied to CAP V2 plotted against the response to the same stimulus applied to CON V2. The solid black line is the curve fit to the data. The hashed line is the ideal relationship for matched perceptual intensity (slope = 1). Mean±SD. B) Post-scan retrospective ratings from the Gracely Pain Scale. The ascending qualitative descriptors were converted to unitary numerical values from 1-15. Each subject made only one report per stimulus condition.

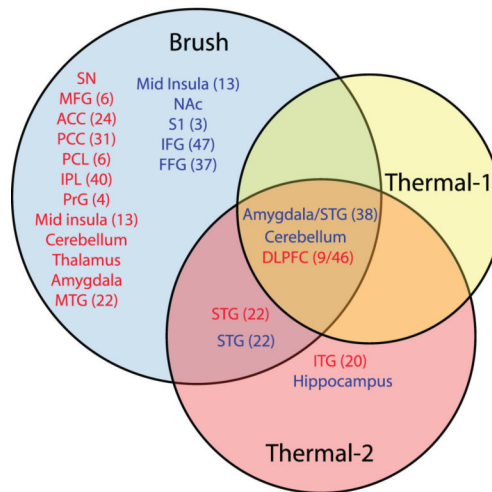


**Figure 5.**  
**Qualitative Psychophysical Reports.** Histogram of sensory and affective descriptors reported for each scan after completion of the scan sessions. Subjects selected one descriptor which best characterized their perceptual experience. The figure is a histogram of those responses across all 12 subjects. Every subject accounted for exactly 1 report for each stimulus condition. The categories along the x-axis are the qualitative descriptors listed in the Gracely Pain Scale.

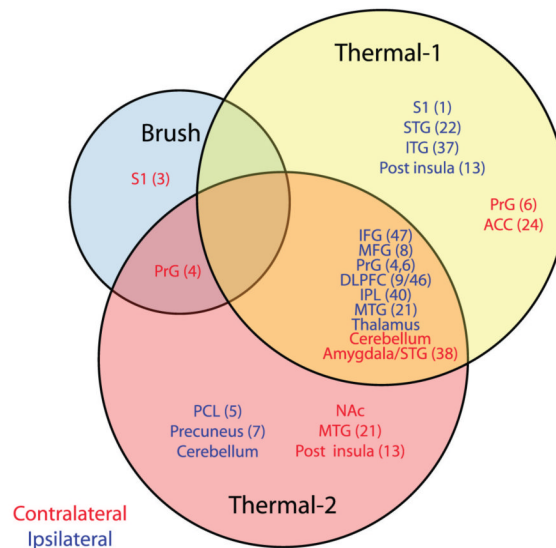
**(A) BOLD responses - Single trial averages for contralateral S1****(B) Direction of median change in ROI sampled COPEs with capsaicin****Figure 6.****Differences in Capsaicin vs. Control V2 responses in trigeminal nociceptive pathway ROIs.**

A) Example of functional time series group single trial average (STA) responses to Brush, Thermal-1, and Thermal-2 stimuli. Black bar denotes application of stimuli. The STA response was shifted in the y-axis for each stimulus condition, such that the median of the five pre-stimulus time points was set to zero. Mean $\pm$ SE. B) Direction of median differences of sampled COPE values from trigeminal ROIs in CAP V2 with respect to CON V2. The S1 and Thal ROIs are contralateral to the stimulus application, while the BS(p), BS(c), and TG ROIs are ipsilateral. All ROIs were found to be important for discrimination in the logistic regression analysis, except for S1 with Thermal-1 stimulation (Supplemental Table 2). Quantitative values are not comparable across ROIs, due to the variable relationship between the BOLD signal and physiological activity across the ROIs. The overall classification accuracy using all ROIs was in the range of 60-70% for all three stimuli.

**Stimulus-dependent laterality contrast**  
**(A) Capsaicin > Control**



**(B) Control > Capsaicin**

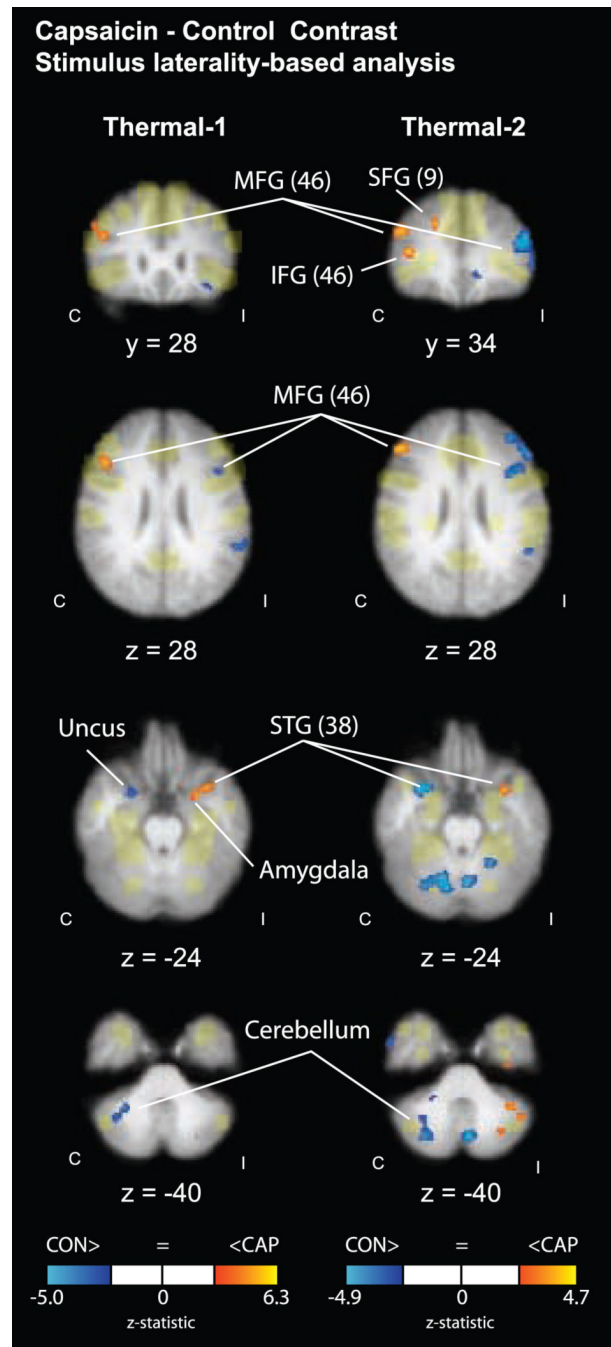


**Figure 7.**

**Venn diagram summary of Capsaicin vs. Control contrast.** (A) Regions that show greater activation in response to stimulation to CAP V2 than to CON V2 (Capsaicin > Control). Capsaicin V2 responses were significantly greater than control in more areas following brush than thermal stimuli. All three stimuli showed increased responses in amygdala, cerebellum, and DLPFC with Capsaicin V2 stimulation. (B) Regions that show greater activation in response to stimulation to CON V2 than to CAP V2 (Capsaicin < Control). Capsaicin V2 responses were significantly lower than control in more areas with thermal stimuli than brush. Red font=areas active contralateral to stimulus. Blue font=areas active ipsilateral to stimulus. S1=Primary somatosensory cortex, MFG=Middle frontal gyrus, DLPFC=Dorsolateral

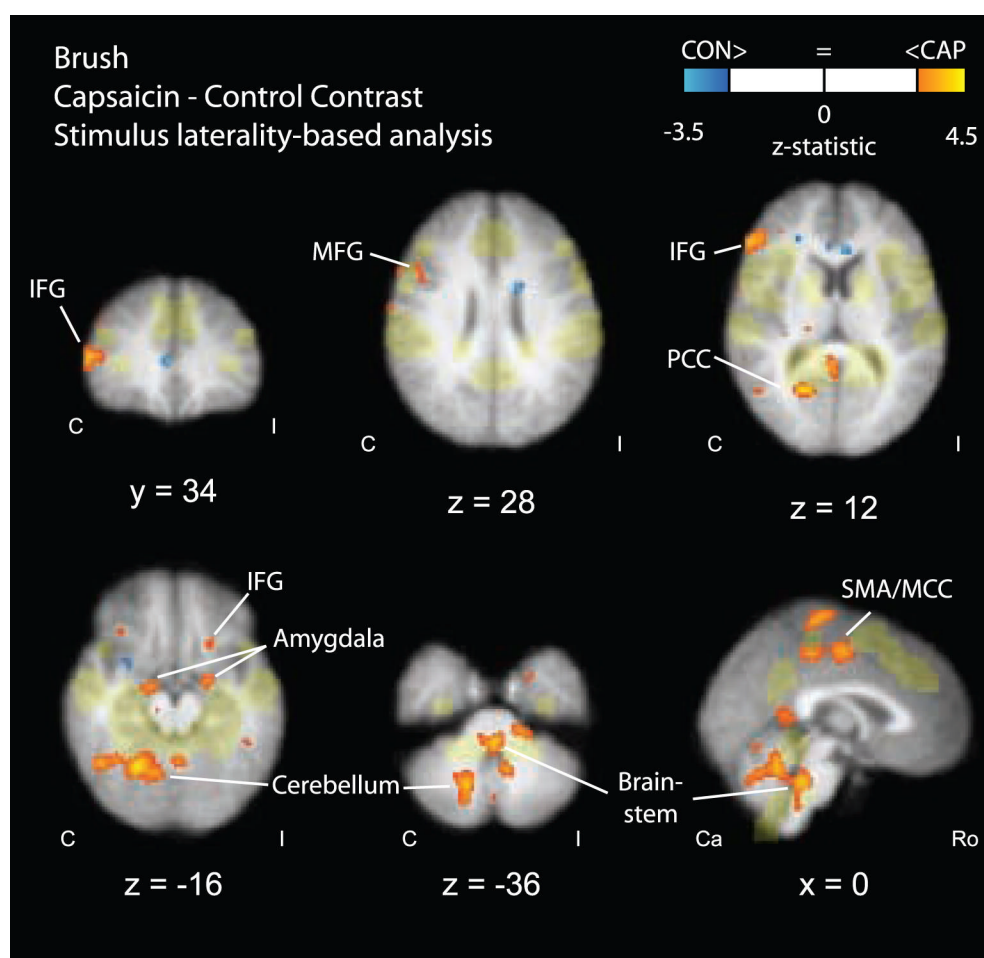
prefrontal cortex, IFG=Inferior frontal gyrus, PrG=Precentral gyrus, ACC=Anterior cingulate cortex, PCC=Posterior cingulate cortex, PCL=Paracentral lobule, IPL=Inferior parietal lobule, SN=Substantia nigra, STG=Superior temporal gyrus, FFG=Fusiform gyrus, MTG=Middle temporal gyrus, ITG=Inferior temporal gyrus, NAc=Nucleus accumbens.





**Figure 8.**

**Capsaicin vs. Control contrast for thermal stimuli.** Transparent yellow areas reflect the hemispheric lateralized activation mask specific for each stimulus (see Methods). Analysis was restricted to areas outside of these masks. Contrast thresholding was performed using generalized mixture modeling (see Methods). For presentation, statistical images have been spatially smoothed using low pass Gaussian linear filtering with a sigma of 2 mm. IFG=inferior frontal gyrus, MFG=middle frontal gyrus, SFG=superior frontal gyrus, STG=superior temporal gyrus.



**Figure 9.**

**Capsaicin vs. Control contrast for brush stimuli.** Transparent yellow areas reflect the hemispheric lateralized activation mask specific for each stimulus. Analysis was restricted to areas outside of these masks. For presentation, statistical images have been spatially smoothed using low pass Gaussian linear filtering with a sigma of 2 mm and an acceptance threshold of 50%. IFG=inferior frontal gyrus, MFG=middle frontal gyrus, SMA=supplementary motor area, MCC=midcingulate cortex, PCC=posterior cingulate cortex.

Table 1A

Contrast analysis – CAP>CON

Area	Side	x	y	z	Max Z
<b>Thermal-1 (<math>z&gt;2.71</math>)</b>					
MFG (9)	C	36	14	32	3.80
Amygdala	I	-24	2	-24	3.24
STG (38)	I	-38	8	-22	3.29
Cerebellum	I	-28	-78	-48	3.81
<b>Thermal-2 (<math>z&gt;2.43</math>)</b>					
SFG (9)	C	20	34	34	3.53
MFG (46)	C	44	34	30	3.27
IFG (46)	C	42	38	14	3.80
STG (22)	C	58	-34	4	3.95
STG (38)	I	-30	8	-24	3.69
Hippocampus	I	-32	-18	-36	3.64
ITG (20)	C	44	-2	-44	4.66
Cerebellum	I	-46	-58	-44	3.44
	I	-32	-68	-38	3.34
<b>Brush (<math>z&gt;3.10</math>)</b>					
MFG (6)	C	22	10	68	3.67
MFG (9)	C	52	20	30	3.24
IFG (46)	C	54	34	10	3.54
IFG (47)	I	-22	22	-16	2.94
Mid. insula (13)	C	44	8	-8	3.43
ACC (24)	I	-36	-6	6	3.11
PCC (31)	C	2	-10	46	3.29
PCL (6)	C	20	-64	14	4.49
IPL (40)	C	12	-26	48	3.79
SI (3)	I	36	-36	42	3.51
PrG (4)	I	-50	-16	50	3.41
Amygdala	C	34	-24	54	3.56
	C	16	-6	-16	3.59
NAC	I	-24	-2	-16	3.01
Thalamus, Pulv	I	6	4	-8	3.22
SN	C	20	-26	8	2.97
STG (22)	C	12	-20	-14	3.10
	C	50	-26	-4	3.34
	I	-56	-8	4	3.54
	I	-46	-52	22	3.29
MTG (22)	C	60	-34	2	4.04
FFG (37)	I	-42	-40	-20	3.50
Cerebellum	C	20	-76	-34	4.54
	I	-50	-60	-42	3.79
	I	-24	-68	-22	3.35
	I	-22	-30	-36	3.52
	I	-24	-68	-22	3.35
	I	-14	-44	-48	3.18

Table 1B. Contrast analysis – CAP<CON

Area	Side	x	y	z	Max Z
<b>Thermal-1 (<math>z&gt;2.83</math>)</b>					

Table 1B. Contrast analysis – CAP&lt;CON

Area	Side	x	Local maxima y	z	Max Z
MFG (8)	I	-32	22	46	3.64
IFG (46)	I	-40	16	24	3.46
IFG (47)	I	-38	26	-16	3.52
Post. insula (13)	I	-46	-22	22	3.57
ACC (24)	C	8	-2	46	3.65
IPL (40)	I	-64	-40	22	3.46
SI	I	-44	-26	52	3.83
PrG (4)	I	-40	-12	52	3.99
PrG (6)	I	-50	-8	40	4.02
	I	-44	-18	64	3.72
Uncus (34)	C	28	4	-26	3.44
Thalamus, MD	I	-8	-16	10	3.26
Thalamus, VL	I	-12	-14	8	3.63
STG (22)	I	-64	-36	12	3.61
MTG (21)	I	-58	-52	0	3.67
	I	-48	-30	-6	3.47
ITG (37)	I	-52	-62	-14	3.90
Cerebellum	C	40	-72	-26	4.10
	C	26	-74	-50	3.56
	C	6	-62	-58	3.43
<b>Thermal-2 (z&gt;3.09)</b>					
SFG (9)	I	-18	50	38	3.52
MFG (8)	I	-24	20	50	3.25
MFG (46)	I	-44	34	22	3.94
IFG (47)	I	-40	22	-16	3.31
Post. insula (13)	C	44	-20	12	3.19
PCL (5)	I	-4	-36	50	3.64
IPL (40)	I	-44	-48	42	4.06
Precuneus (7)	I	-14	-44	46	3.72
PrG (4)	C	54	-14	58	3.77
	C	26	-30	56	3.86
	I	-52	-14	42	3.57
PrG (6)	I	-62	-6	12	3.50
Thalamus, MD	I	-8	-16	14	3.60
Thalamus, VL	I	-16	-12	8	3.09
STG (38)	C	32	8	-20	4.22
MTG (21)	I	-54	-60	0	3.73
	I	-50	-26	-6	4.05
NAc	C	56	4	-36	3.25
Cerebellum	C	14	2	-2	2.85
	C	38	-72	-28	4.56
	C	8	-62	-44	3.34
	I	-8	-64	-14	3.94
	I	-4	-76	-40	4.07
	I	-22	-52	-24	3.44
	I	-26	-58	-60	4.08
<b>Brush (z&gt;2.67)</b>					
SI (3)	C	48	-20	58	3.28
PrG (4)	C	38	-18	60	3.37
	C	48	-20	42	3.34

**Key:** Thresholds were determined using the Gaussian Mixture Modeling technique described in the text. SI=Primary somatosensory cortex, SFG=Superior frontal gyrus, MFG=Middle frontal gyrus, IFG=Inferior frontal gyrus, STG=Superior temporal gyrus, MTG=Middle temporal gyrus, ITG=Inferior temporal gyrus, ACC=Anterior cingulate cortex, PCC=Posterior cingulate cortex.

PCL=Paracentral lobule, IPL=Inferior parietal lobule, PrG=Precentral gyrus, NAc=Nucleus accumbens, Pulv=Pulvinar, SN=Substantia nigra, FFG=Fusiform gyrus, MD=Mediodorsal thalamic nucleus, VL=Ventrolateral thalamic nucleus.