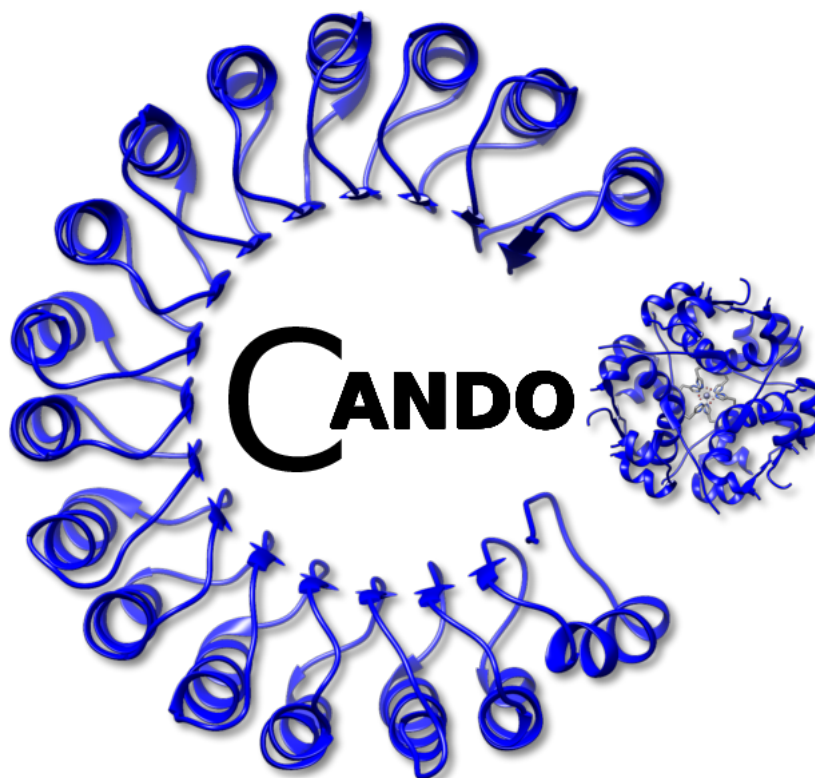

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1 CANDO

Computational **A**nalysis of **N**ovel **D**rug **O**pportunities

CANDO is a unique computational drug discovery, design, and repurposing platform.

2 Install

You may download the source code via the releases or cloning the git repository. However, we suggest using anaconda to install the CANDO package, as this is the easiest and quickest way to start using our platform!

The CANDO package relies on multiple "conda-forge" dependencies. Therefore, we require that you add "conda-forge" to your anaconda channels:

```
conda config --add channels conda-forge
```

Then you can install CANDO using the following command:

```
conda install -c ram-compbio cando
```

3 Test

You can test your install by running our script:

`test.py`

4 Authors

- William Mangione
- Zackary Falls
- James Schuler
- Matt Hudson
- Liana Bruggemann
- Ram Samudrala

For general questions, please contact Ram Samudrala (ram@compbio.org). For direct questions about source code for [cando.py](#), please contact William Mangione (wmangion@buffalo.edu) or Zackary Falls (zmfalls@buffalo.edu).

5 LICENSE

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6 Namespace Index

6.1 Packages

Here are the packages with brief descriptions (if available):

cando	4
-----------------------	---

7 Hierarchical Index

7.1 Class Hierarchy

This inheritance list is sorted roughly, but not completely, alphabetically:

object	
cando.ADR	10
cando.CANDO	11
cando.Compound	29
cando.Indication	32
cando.Matrix	33
cando.Pathway	36
cando.Protein	37

8 Class Index

8.1 Class List

Here are the classes, structs, unions and interfaces with brief descriptions:

cando.ADR		10
An object to represent an adverse reaction		
cando.CANDO		11
An object to represent all aspects of CANDO (compounds, indications, matrix, etc.)		
cando.Compound		29
An object to represent a compound/drug		
cando.Indication		32
An object to represent an indication (disease)		
cando.Matrix		33
An object to represent a matrix		

cando.Pathway	
An object to represent a pathway	36
cando.Protein	
An object to represent a protein	37

9 File Index

9.1 File List

Here is a list of all files with brief descriptions:

cando.py	38
--------------------------	----

10 Namespace Documentation

10.1 cando Namespace Reference

Classes

- class [ADR](#)
An object to represent an adverse reaction.
- class [CANDO](#)
An object to represent all aspects of [CANDO](#) (compounds, indications, matrix, etc.)
- class [Compound](#)
An object to represent a compound/drug.
- class [Indication](#)
An object to represent an indication (disease)
- class [Matrix](#)
An object to represent a matrix.
- class [Pathway](#)
An object to represent a pathway.
- class [Protein](#)
An object to represent a protein.

Functions

- def [generate_matrix](#) (cmpd_scores, prot_scores, c_cutoff=0.0, p_cutoff=0.0, percentile_cutoff=None, interaction_score='P', matrix_file='cando_interaction_matrix.tsv', ncpus=1)
Generate a [CANDO Matrix](#).
- def [generate_scores](#) (fp="rd_ecfp4", cmpd_pdb="", out_path='.')
Generate the fingerprint for a new compound and calculate the Tanimoto similarities against all binding site ligands.
- def [generate_signature](#) (cmpd_scores="", prot_scores="", c_cutoff=0.0, p_cutoff=0.0, percentile_cutoff=None, interaction_score='P', matrix_file="")
Generate signature.
- def [get_scores](#) (c, p_scores, c_score, c_cutoff, p_cutoff, percentile_cutoff, i_score)
Get best score for each Compound-Protein interaction.

- def [score_fp](#) (fp, compd_file, compd_id, bs)
Generate the scores for a given [Compound](#) against all [Protein](#) ligands.
- def [cosine_dist](#) (A)
- def [tanimoto_sparse](#) (str1, str2)
Calculate the tanimoto coefficient for a pair of sparse vectors.
- def [tanimoto_dense](#) (list1, list2)
Calculate the tanimoto coefficient for a pair of dense vectors.
- def [get_fp_lig](#) (fp)
Download precompiled binding site ligand fingerprints using the given fingerprint method.
- def [get_v2](#) (matrix='nrpdb')
Download [CANDO](#) v2.0 data.
- def [get_tutorial](#) ()
Download data for tutorial.
- def [get_test](#) ()
Download data for test script.
- def [dl_dir](#) (url, out, l)
Function to recursively download a directory.
- def [dl_file](#) (url, out_file)
Function to download a file.

10.1.1 Function Documentation

10.1.1.1 [cosine_dist\(\)](#) `def cando.cosine_dist (`
 `A)`

10.1.1.2 [dl_dir\(\)](#) `def cando.dl_dir (`
 `url,`
 `out,`
 `l)`

Function to recursively download a directory.

Prints the name of the directory and a progress bar.

Parameters

<i>url</i>	str: URL of the dir to be downloaded
<i>out</i>	str: Path to where the dir will be downloaded
<i>l</i>	list: List of files in dir to be downloaded

10.1.1.3 [dl_file\(\)](#) `def cando.dl_file (`
 `url,`
 `out_file)`

Function to download a file.

Prints the name of the file and a progress bar.

Parameters

<i>url</i>	str: URL of the file to be downloaded
<i>out_file</i>	str: File path to where the file will be downloaded

10.1.1.4 generate_matrix() `def cando.generate_matrix (`
 cmpd_scores,
 prot_scores,
 c_cutoff = 0.0,
 p_cutoff = 0.0,
 percentile_cutoff = None,
 interaction_score = 'P',
 matrix_file = 'cando_interaction_matrix.tsv',
 ncpus = 1)

Generate a [CANDO Matrix](#).

Parameters

<i>cmpd_scores</i>	str: File path to tab-separated scores for all Compounds
<i>prot_scores</i>	str: File path to tab-separated scores for all Proteins
<i>c_cutoff</i>	Any Cscores below this value will not be considered for the interaction score. (0.0-1.0). Default = 0.0
<i>p_cutoff</i>	Any Pcores below this value will not be considered for the interaction score. (0.0-1.0). Default = 0.0
<i>percentile_cutoff</i>	Percentile of all Compound-ligand Cscores for each Compound by which the Cscore cutoff will be defined (0.0-100.0 or None). This makes the hard <i>c_cutoff</i> variable for each Compound to avoid molecular size bias due to fingerprinting. This overwrites the use of <i>c_cutoff</i> . Default = None
<i>interaction_score</i>	The scoring function for the interaction between each Compound-Protein pair. ('C', 'dC', 'P', 'CxP', 'dCxP').
<i>matrix_file</i>	str: File path to where the generated Matrix will be written Default = 'cando_interaction_matrix.tsv'
<i>ncpus</i>	int: Number of cpus to use for parallelization. Default = 1

10.1.1.5 generate_scores() `def cando.generate_scores (`
 fp = "rd_ecfp4",
 cmpd_pdb = '',
 out_path = '.')

Generate the fingerprint for a new compound and calculate the Tanimoto similarities against all binding site ligands.

Parameters

<i>fp</i>	str: The fingerprinting software and method used, e.g. 'rd_ecfp4', 'ob_fp2'
<i>compd_pdb</i>	str: File path to the PDB
<i>out_path</i>	str: Path to where the scores file will be written

10.1.1.6 generate_signature() `def cando.generate_signature (`
 compd_scores = '',
 prot_scores = '',
 c_cutoff = 0.0,
 p_cutoff = 0.0,
 percentile_cutoff = None,
 interaction_score = 'P',
 matrix_file = '')

Generate signature.

Parameters

<i>compd_scores</i>	str: File path to tab-separated scores for all Compounds
<i>prot_scores</i>	str: File path to tab-separated scores for all Proteins
<i>matrix_file</i>	str: File path to where the generated Compounds signature will be written
<i>c_cutoff</i>	Any Cscores below this value will not be considered for the interaction score. (0.0-1.0). Default = 0.0
<i>p_cutoff</i>	Any Pcores below this value will not be considered for the interaction score. (0.0-1.0). Default = 0.0
<i>percentile_cutoff</i>	Percentile of all Compound-ligand Cscores for each Compound by which the Cscore cutoff will be defined (0.0-100.0 or None). This makes the hard <i>c_cutoff</i> variable for each Compound to avoid molecular size bias due to fingerprinting. This overwrites the use of <i>c_cutoff</i> . Default = None
<i>interaction_score</i>	The scoring function for the interaction between each Compound-Protein pair. ('C', 'dC', 'P', 'CxP', 'dCxP').

10.1.1.7 get_fp_lig() `def cando.get_fp_lig (`
 fp)

Download precompiled binding site ligand fingerprints using the given fingerprint method.

Parameters

<i>fp</i>	str: Fingerprinting method used to compile each binding site ligand fingerprint
-----------	---

10.1.1.8 get_scores() `def cando.get_scores (`

```

c,
p_scores,
c_score,
c_cutoff,
p_cutoff,
percentile_cutoff,
i_score )

```

Get best score for each Compound-Protein interaction.

Parameters

<i>c</i>	int: Compound id
<i>p_scores</i>	df: DataFrame of all Protein ligands and corresponding scores
<i>c_score</i>	df: DataFrame of all Compound-ligand scores
<i>c_cutoff</i>	Any Cscores below this value will not be considered for the interaction score. (0.0-1.0).
<i>p_cutoff</i>	Any Pcores below this value will not be considered for the interaction score. (0.0-1.0).
<i>percentile_cutoff</i>	Percentile of all Compound-ligand Cscores for each Compound by which the Cscore cutoff will be defined (0.0-100.0). This makes the hard c_cutoff variable for each Compound to avoid molecular size bias due to fingerprinting. This overwrites the use of c_cutoff.
<i>i_score</i>	The scoring function for the interaction between each Compound-Protein pair. (C, dC, P, CxP, dCxP).

10.1.1.9 `get_test()` `def cando.get_test ()`

Download data for test script.

This data includes:

- Test [Matrix](#) (Approved drugs (2,162) and 64 proteins)
- v2.0 [Compound](#) mapping (approved and all)
- v2.0 [Indication](#) - [Compound](#) mapping
- [Compound](#) scores file for all approved compounds (fingerprint: rd_ecfp4)
- Test [Protein](#) scores file (64 proteins) for all binding site ligands for each [Protein](#) (fingerprint: rd_ecfp4)
- Test [Compound](#) in PDB format to generate a new fingerprint and vector in the [Matrix](#)
- Directory of test Compounds in PDB format to generate multiple new fingerprints and vectors in the [Matrix](#)
- Test Pathways set

10.1.1.10 get_tutorial() `def cando.get_tutorial ()`

Download data for tutorial.

This data includes:

- Example [Matrix](#) (Approved drugs (2,162) and 64 proteins)
- v2.0 [Compound](#) mapping (approved and all)
- v2.0 [Indication](#) - [Compound](#) mapping
- [Compound](#) scores file for all approved compounds (fingerprint: rd_ecfp4)
- Example [Protein](#) scores file (64 proteins) for all binding site ligands for each [Protein](#) (fingerprint: rd_ecfp4)
- Example [Compound](#) in PDB format to generate a new fingerprint and vector in the [Matrix](#)
- Example Pathways set

10.1.1.11 get_v2() `def cando.get_v2 (
 matrix = 'nrpdb')`

Download [CANDO](#) v2.0 data.

This data includes:

- [Compound](#) mapping (approved and all)
- Indication-compound mapping
- Scores file for all approved compounds (fingerprint: rd_ecfp4)
- [Matrix](#) file for approved drugs (2,162) and all proteins (14,610) (fingerprint: rd_ecfp4)

10.1.1.12 score_fp() `def cando.score_fp (
 fp,
 compd_file,
 compd_id,
 bs)`

Generate the scores for a given [Compound](#) against all [Protein](#) ligands.

Parameters

<i>fp</i>	str: Fingerprinting software and method used, e.g., rd_ecfp4
<i>compd_file</i>	str: File path to PDB
<i>compd_id</i>	int: Number corresponding to the new Compound id
<i>bs</i>	df: DataFrame of all protein ligand fingerprints for the given fingerprinting method (fp)

10.1.1.13 `tanimoto_dense()` `def cando.tanimoto_dense (`
 `list1,`
 `list2)`

Calculate the tanimoto coefficient for a pair of dense vectors.

Parameters

<i>list1</i>	list: List of positions that have a 1 in first compound fingerprint
<i>list2</i>	list: List of positions that have a 1 in second compound fingerprint

10.1.1.14 `tanimoto_sparse()` `def cando.tanimoto_sparse (`
 `str1,`
 `str2)`

Calculate the tanimoto coefficient for a pair of sparse vectors.

Parameters

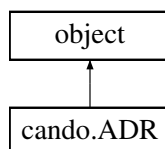
<i>str1</i>	str: String of 1s and 0s representing the first compound fingerprint
<i>str2</i>	str: String of 1s and 0s representing the second compound fingerprint

11 Class Documentation

11.1 `cando.ADR` Class Reference

An object to represent an adverse reaction.

Inheritance diagram for `cando.ADR`:



Public Member Functions

- `def __init__ (self, id_, name)`

Public Attributes

- [id_](#)
- [name](#)
- [compounds](#)

11.1.1 Detailed Description

An object to represent an adverse reaction.

11.1.2 Constructor & Destructor Documentation

11.1.2.1 `__init__()` `def cando.ADR.__init__ (`
 `self,`
 `id_,`
 `name)`

11.1.3 Member Data Documentation

11.1.3.1 `compounds` `cando.ADR.compounds`

list: [Compound](#) objects associated with the given [ADR](#)

11.1.3.2 `id_` `cando.ADR.id_`

str: Identification for the given [ADR](#)

11.1.3.3 `name` `cando.ADR.name`

str: Name of the given [ADR](#)

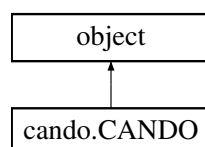
The documentation for this class was generated from the following file:

- [cando.py](#)

11.2 cando.CANDO Class Reference

An object to represent all aspects of [CANDO](#) (compounds, indications, matrix, etc.)

Inheritance diagram for `cando.CANDO`:



Public Member Functions

- def `__init__` (self, `c_map`, `i_map`, `matrix`="", `compute_distance`=False, `save_rmsds`="", `read_rmsds`="", `pathways`="", `pathway_quantifier`='max', `indication_pathways`="", `indication_proteins`="", `similarity`=False, `dist_metric`='rmsd', `protein_set`="", `rm_zeros`=False, `rm_compounds`="", `adr_map`="", `protein_map`="", `nopus`=1)
- def `search_compound` (self, name, n=5)
Print closest [Compound](#) names/IDs for input search str.
- def `get_compound` (self, `compd_id`)
Get [Compound](#) object from [Compound](#) id or fuzzy match to [Compound](#) name.
- def `get_protein` (self, `protein_id`)
Get [Protein](#) object from [Protein](#) id.
- def `get_indication` (self, `ind_id`)
Get [Indication](#) object from [Indication](#) id.
- def `get_pathway` (self, `id_`)
Get [Pathway](#) object from [Pathway](#) id.
- def `get_adr` (self, `id_`)
Get [ADR](#) (adverse drug reaction) from [ADR](#) id.
- def `search_indication` (self, name, n=5)
Print closest MeSH IDs for [Indication](#) name.
- def `top_targets` (self, `compd`, n=10, `negative`=False)
Get the top scoring protein targets for a given compound.
- def `common_targets` (self, `compds_file`, n=10, `negative`=False, `save_file`="")
Get the top scoring protein targets for a given compound.
- def `virtual_screen` (self, `protein`, n=10, `negative`=False, `compound_set`='all', `save_file`="")
Get the top scoring protein targets for a given compound.
- def `uniprot_set_index` (self, `prots`)
Gather proteins from input matrix that map to UniProt IDs from 'protein_set=' param.
- def `generate_similar_sigs` (self, `compd`, `sort`=False, `proteins`=[], `aux`=False)
For a given compound, generate the similar compounds using distance of sigs.
- def `generate_some_similar_sigs` (self, `compds`, `sort`=False, `proteins`=[], `aux`=False)
For a given list of compounds, generate the similar compounds based on rmsd of sigs This is pathways/genes for all intents and purposes.
- def `quantify_pathways` (self, `indication`=None)
Uses the pathway quantifier defined in the [CANDO](#) instantiation to make a pathway signature for all pathways in the input file (NOTE: does not compute distances)
- def `results_analysed` (self, `f`, `metrics`, `effect_type`)
Creates the results analysed named file for the benchmarking and computes final avg indication accuracies.
- def `canbenchmark` (self, `file_name`, `indications`=[], `continuous`=False, `bottom`=False, `ranking`='standard', `adrs`=False)
Benchmarks the platform based on compound similarity of those approved for the same diseases.
- def `canbenchmark_associated` (self, `file_name`, `indications`=[], `continuous`=False, `ranking`='standard')
Benchmark only the compounds in the indication mapping, aka get rid of "noisy" compounds.
- def `canbenchmark_bottom` (self, `file_name`, `indications`=[], `ranking`='standard')
Benchmark the reverse ranking of similar compounds as a control.
- def `canbenchmark_ndcg` (self, `file_name`)
- def `canbenchmark_cluster` (self, `n_clusters`=5)
Benchmark using k-means clustering.
- def `ml` (self, `method`='rf', `effect`=None, `benchmark`=False, `adrs`=False, `predict`=[], `threshold`=0.5, `negative`='random', `seed`=42, `out`="")
create an ML classifier for a specified indication or all inds (to benchmark) predict (used w/ 'effect=' - indication or [ADR](#)) is a list of compounds to classify with the trained ML model out=X saves benchmark SUMMARY->SUMMARY<-Y_ml_X; raw results->raw_results/raw_results_ml_X (same for RAN) currently supports random forest ('rf'), support vector machine ('svm'), 1-class SVM ('1csvm'), and logistic regression ('log') models are trained with leave-one-out cross validation during benchmarking

- def [raw_results_roc](#) (self, rr_files, labels, save='roc-raw_results.pdf')
- def [canpredict_denovo](#) (self, method='count', threshold=0.0, topX=10, ind_id=None, [proteins](#)=None, [compd_set](#)='all', save='')
This function is used for predicting putative therapeutics for an indication of interest by summing/counting the number of interactions above a certain input interaction threshold for all proteins or a specified subset of proteins.
- def [canpredict_compounds](#) (self, ind_id, n=10, topX=10, keep_associated=False, [compd_set](#)='all', save='')
This function is used for predicting putative therapeutics for an indication of interest using a homology-based approach.
- def [canpredict_indications](#) (self, compd, n=10, topX=10, save='')
This function is the inverse of canpredict_compounds.
- def [similar_compounds](#) (self, compd, n=10)
Computes and prints the top n most similar compounds to an input [Compound](#) object cando_compd or input novel signature new_sig.
- def [add_compd](#) (self, new_sig, new_name='')
Add a new [Compound](#) object to the platform.
- def [sigs](#) (self, rm)
Return a list of all signatures, rm is a list of compound ids you do not want in the list.
- def [save_rmsds_to_file](#) (self, f)
Write calculated distances of all compounds to all compounds to file.
- def [fusion](#) (self, cando_objs, out_file="", method='sum')
This function re-ranks the compounds according to the desired comparison specified by 'method' -> currently supports 'min', 'avg', 'mult', and 'sum'.
- def [normalize](#) (self)
Normalize the distance scores to between [0,1].
- def [__str__](#) (self)
Print stats about the [CANDO](#) object.

Public Attributes

- [c_map](#)
- [i_map](#)
- [matrix](#)
- [protein_set](#)
- [pathways](#)
- [accuracies](#)
- [compute_distance](#)
- [clusters](#)
- [rm_zeros](#)
- [rm_compounds](#)
- [rm_cmpds](#)
- [save_rmsds](#)
- [read_rmsds](#)
- [similarity](#)
- [dist_metric](#)
- [ncpus](#)
- [pathway_quantifier](#)
- [indication_pathways](#)
- [indication_proteins](#)
- [adr_map](#)
- [protein_map](#)
- [proteins](#)
- [protein_id_to_index](#)
- [compounds](#)

- [indications](#)
- [indication_ids](#)
- [adrs](#)
- [short_matrix_path](#)
- [short_read_rmsds](#)
- [short_protein_set](#)
- [cmpd_set](#)
- [data_name](#)

11.2.1 Detailed Description

An object to represent all aspects of [CANDO](#) (compounds, indications, matrix, etc.)

To instantiate you need the compound mapping (`c_map`), an indication mapping file (`i_map`), and typically and a compound-protein matrix (`matrix=`) or or precomputed compound-compound distance matrix (`read_rmsds=`), but those are optional.

11.2.2 Constructor & Destructor Documentation

11.2.2.1 `__init__()` `def cando.CANDO.__init__ (`
 `self,`
 `c_map,`
 `i_map,`
 `matrix = '',`
 `compute_distance = False,`
 `save_rmsds = '',`
 `read_rmsds = '',`
 `pathways = '',`
 `pathway_quantifier = 'max',`
 `indication_pathways = '',`
 `indication_proteins = '',`
 `similarity = False,`
 `dist_metric = 'rmsd',`
 `protein_set = '',`
 `rm_zeros = False,`
 `rm_compounds = '',`
 `adr_map = '',`
 `protein_map = '',`
 `n_cpus = 1)`

11.2.3 Member Function Documentation

11.2.3.1 `__str__()` `def cando.CANDO.__str__ (`
 `self)`

Print stats about the [CANDO](#) object.

11.2.3.2 add_cmpd() `def cando.CANDO.add_cmpd (`
 `self,`
 `new_sig,`
 `new_name = '')`

Add a new [Compound](#) object to the platform.

Parameters

<i>new_sig</i>	str: Path to the tab-separated interaction scores
<i>new_name</i>	str: Name for the new Compound

Returns

cmpd [Compound](#): [Compound](#) object

11.2.3.3 canbenchmark() `def cando.CANDO.canbenchmark (`
 `self,`
 `file_name,`
 `indications = [],`
 `continuous = False,`
 `bottom = False,`
 `ranking = 'standard',`
 `adrs = False)`

Benchmarks the platform based on compound similarity of those approved for the same diseases.

Parameters

<i>file_name</i>	str: Name to be used for the various results files (e.g. file_name=test --> summary_test.tsv)
<i>indications</i>	list or str: List of Indication ids to be used for this benchmark, otherwise all will be used.
<i>continuous</i>	bool: Use the percentile of distances from the similarity matrix as the cutoffs for benchmarking
<i>bottom</i>	bool: Reverse the ranking (descending) for the benchmark
<i>ranking</i>	str: What ranking method to use for the compounds. This really only affects ties. (standard, modified, and ordinal)
<i>adrs</i>	bool: ADRs are used as the Compounds' phenotypic effects instead of Indications

11.2.3.4 canbenchmark_associated() `def cando.CANDO.canbenchmark_associated (`
 `self,`
 `file_name,`
 `indications = [],`
 `continuous = False,`
 `ranking = 'standard')`

Benchmark only the compounds in the indication mapping, aka get rid of "noisy" compounds.

This function returns the filtered [CANDO](#) object in the event that you want to explore further.

Parameters

<i>file_name</i>	str: Name to be used for the various results files (e.g. file_name=test --> summary_test.tsv)
<i>indications</i>	list: List of Indication ids to be used for this benchmark, otherwise all will be used.
<i>continuous</i>	bool: Use the percentile of distances from the similarity matrix as the cutoffs for benchmarking
<i>ranking</i>	str: What ranking method to use for the compounds. This really only affects ties. (standard, modified, and ordinal)

11.2.3.5 canbenchmark_bottom() `def cando.CANDO.canbenchmark_bottom (`
 self,
 file_name,
 indications = [],
 ranking = 'standard')

Benchmark the reverse ranking of similar compounds as a control.

Parameters

<i>file_name</i>	str: Name to be used for the various results files (e.g. file_name=test --> summary_test.tsv)
<i>indications</i>	list: List of Indication ids to be used for this benchmark, otherwise all will be used.
<i>ranking</i>	str: What ranking method to use for the compounds. This really only affects ties. (standard, modified, and ordinal)

11.2.3.6 canbenchmark_cluster() `def cando.CANDO.canbenchmark_cluster (`
 self,
 n_clusters = 5)

Benchmark using k-means clustering.

Parameters

<i>n_clusters</i>	int: Number of clusters for k-means
-------------------	-------------------------------------

11.2.3.7 canbenchmark_ndcg() `def cando.CANDO.canbenchmark_ndcg (`
 self,
 file_name)

11.2.3.8 canpredict_compounds() `def cando.CANDO.canpredict_compounds (`
 self,
 ind_id,

```

n = 10,
topX = 10,
keep_associated = False,
cmpd_set = 'all',
save = '' )

```

This function is used for predicting putative therapeutics for an indication of interest using a homology-based approach.

Input an `ind_id` id and for each of the associated compounds, it will generate the similar compounds (based on distance) and add them to a dictionary with a value of how many times it shows up (enrichment). If a compound not approved for the indication of interest keeps showing up, that means it is similar in signature to the drugs that are ALREADY approved for the indication, so it may be a target for repurposing. Control how many similar compounds to consider with the argument '`n`'. In the output, '`score1`' refers to the number of times the compound shows up in the top '`n`' drugs associated with the indication and '`score2`' is the average of the ranks for '`score1`' (note: '`score2`' <= '`n`').

Parameters

<code>ind_id</code>	str: Indication id
<code>n</code>	int: top number of similar Compounds to be used for each Compound associated with the given Indication
<code>topX</code>	int: top number of predicted Compounds to be printed
<code>keep_associated</code>	bool: Print Compounds that are already approved/associated for the Indication
<code>cmpd_set</code>	str: specify the compound set to use ('all', 'approved', or 'other')
<code>save</code>	str: name of a file to save results

```

11.2.3.9 canpredict_denovo() def cando.CANDO.canpredict_denovo (
    self,
    method = 'count',
    threshold = 0.0,
    topX = 10,
    ind_id = None,
    proteins = None,
    cmpd_set = 'all',
    save = '' )

```

This function is used for predicting putative therapeutics for an indication of interest by summing/counting the number of interactions above a certain input interaction threshold for all proteins or a specified subset of proteins.

An indication can be specified to mark drugs associated with that indication in the output. The threshold will vary based on the values of the input matrix. Method can either be '`count`' (`score1`), which ranks compounds based on the number of interactions above the threshold, or '`sum`' (`score2`), which ranks the compounds based on the highest total sum for interaction scores above the threshold (these two are highly correlated but can differ for larger sets of proteins or lower thresholds). A third option is '`targets`', which inspects and outputs the top protein interactions on an individual basis without summing/counting per drug (the output format differs from the other two options). If `indication_proteins` flag is used for the [CANDO](#) object instantiation, the proteins associated with the input indication will automatically be used. Otherwise, the '`proteins=`' input can be used. The output can be saved to a file specified by '`save=`'. If `ind_id` is used, compounds associated with the indication will be included and marked in the output for comparison.

Parameters

<i>method</i>	str: 'sum', 'count', or 'targets'
<i>threshold</i>	float: a interaction score cutoff to use (ignores values for sum/count less than threshold)
<i>topX</i>	int: top number of predicted Compounds to be printed/saved
<i>ind_id</i>	str: an indication id for marking drug output/ specifying protein set
<i>proteins</i>	List str: list of protein IDs from the matrix to use for the sum/count
<i>cmpd_set</i>	str: specify the compound set to use ('all', 'approved', or 'other')
<i>save</i>	str: name of a file to save results

11.2.3.10 canpredict_indications() `def cando.CANDO.canpredict_indications (`
`self,`
`cmpd,`
`n = 10,`
`topX = 10,`
`save = '')`

This function is the inverse of canpredict_compounds.

Input a compound of interest cando_cmpd (or a novel protein signature of interest new_sig) and the most similar compounds to it will be computed. The indications associated with the top n most similar compounds to the query compound will be examined to see if any are repeatedly enriched.

Parameters

<i>cmpd</i>	Compound: Compound object to be used
<i>n</i>	int: top number of similar Compounds to be used for prediction
<i>topX</i>	int: top number of predicted Indications to be printed

11.2.3.11 common_targets() `def cando.CANDO.common_targets (`
`self,`
`cmpds_file,`
`n = 10,`
`negative = False,`
`save_file = '')`

Get the top scoring protein targets for a given compound.

Parameters

<i>cmpds_file</i>	str: list of Compound IDs for which to search common targets
<i>n</i>	int: number of top targets to print/return
<i>negative</i>	int: if the interaction scores are negative (stronger) energies
<i>save_file</i>	str: save results to file name

Returns

Returns list: list of tuples (protein id_, score)

11.2.3.12 fusion()

```
def cando.CANDO.fusion (
    self,
    cando_objs,
    out_file = '',
    method = 'sum' )
```

This function re-ranks the compounds according to the desired comparison specified by 'method' -> currently supports 'min', 'avg', 'mult', and 'sum'.

Parameters

<i>cando_objs</i>	list: List of CANDO objects
<i>out_file</i>	str: Path to where the result will be written
<i>method</i>	str: Method of fusion to be used (e.g., sum, mult, etc.)

11.2.3.13 generate_similar_sigs()

```
def cando.CANDO.generate_similar_sigs (
    self,
    compd,
    sort = False,
    proteins = [],
    aux = False )
```

For a given compound, generate the similar compounds using distance of sigs.

Parameters

<i>compd</i>	object: Compound object
<i>sort</i>	bool: Sort the list of similar compounds
<i>proteins</i>	list: Protein objects to identify a subset of the Compound signature
<i>aux</i>	bool: Use an auxiliary signature (default: False)

Returns

Returns list: Similar Compounds to the given [Compound](#)

11.2.3.14 generate_some_similar_sigs()

```
def cando.CANDO.generate_some_similar_sigs (
    self,
    compds,
    sort = False,
```

```

    proteins = [],
    aux = False )

```

For a given list of compounds, generate the similar compounds based on rmsd of sigs This is pathways/genes for all intents and purposes.

Parameters

<i>cmpds</i>	list: Compound objects
<i>sort</i>	bool: Sort similar compounds for each Compound
<i>proteins</i>	list: Protein objects to identify a subset of the Compound signature
<i>aux</i>	bool: Use an auxiliary signature (default: False)

Returns

Returns list: Similar Compounds to the given [Compound](#)

11.2.3.15 get_adr()

```
def cando.CANDO.get_adr (
    self,
    id_ )
```

Get [ADR](#) (adverse drug reaction) from [ADR](#) id.

Parameters

<i>id_</i>	str: ADR id
<i>_</i>	

Returns

Returns object: [ADR](#) object

11.2.3.16 get_compound()

```
def cando.CANDO.get_compound (
    self,
    compd_id )
```

Get [Compound](#) object from [Compound](#) id or fuzzy match to [Compound](#) name.

Parameters

<i>compd_id</i>	int or str: Compound id or Compound name
-----------------	--

Returns

Returns object: [Compound](#) object or None if no exact match is found

11.2.3.17 get_indication() `def cando.CANDO.get_indication (`
 `self,`
 `ind_id)`

Get [Indication](#) object from [Indication](#) id.

Parameters

<code>ind↔ _id</code>	str: Indication id
---------------------------	------------------------------------

Returns

Returns object: [Indication](#) object

11.2.3.18 get_pathway() `def cando.CANDO.get_pathway (`
 `self,`
 `id_)`

Get [Pathway](#) object from [Pathway](#) id.

Parameters

<code>id↔ _↔</code>	str: Pathway id
-------------------------	---------------------------------

Returns

Returns object: [Pathway](#) object

11.2.3.19 get_protein() `def cando.CANDO.get_protein (`
 `self,`
 `protein_id)`

Get [Protein](#) object from [Protein](#) id.

Parameters

<code>protein↔ _id</code>	str: Protein name
-------------------------------	-----------------------------------

Returns

Returns object: [Protein](#) object

```
11.2.3.20 ml() def cando.CANDO.ml (
    self,
    method = 'rf',
    effect = None,
    benchmark = False,
    adrs = False,
    predict = [],
    threshold = 0.5,
    negative = 'random',
    seed = 42,
    out = '' )
```

create an ML classifier for a specified indication or all inds (to benchmark) predict (used w/ 'effect=' - indication or [ADR](#)) is a list of compounds to classify with the trained ML model out=X saves benchmark SUMMARY->SUMMARY_ml_X; raw results->raw_results/raw_results_ml_X (same for RAN) currently supports random forest ('rf'), support vector machine ('svm'), 1-class SVM ('1csvm'), and logistic regression ('log') models are trained with leave-one-out cross validation during benchmarking

Parameters

<i>method</i>	str: type of machine learning algorithm to use ('rf', 'svm', '1csvm', and 'log')
<i>effect</i>	Indication or ADR : provide a specific Indication or ADR object to train a classifier
<i>benchmark</i>	bool: benchmark the ML pipeline by training a classifier with LOOCV for each Indication or ADR
<i>adrs</i>	bool: if the models are trained with ADRs instead of Indications
<i>predict</i>	list: provide a list of Compound objects to classify with the model (only used in combination with effect=Indication/ADR object)
<i>threshold</i>	float: decision threshold for positive vs negative classification
<i>negative</i>	str: choose random negative samples (default) or 'inverse' for most opposite signatures
<i>seed</i>	int: choose a seed for reproducibility
<i>out</i>	str: file name extension for the output of benchmark (note: must have benchmark=True)

```
11.2.3.21 normalize() def cando.CANDO.normalize (
    self )
```

Normalize the distance scores to between [0,1].

Simply divides all scores by the largest distance between any two compounds.

```
11.2.3.22 quantify_pathways() def cando.CANDO.quantify_pathways (
    self,
    indication = None )
```

Uses the pathway quantifier defined in the [CANDO](#) instantiation to make a pathway signature for all pathways in the input file (NOTE: does not compute distances)

Parameters

<i>indication</i>	object: Indication object
-------------------	---

11.2.3.23 raw_results_roc() `def cando.CANDO.raw_results_roc (`
 self,
 rr_files,
 labels,
 save = 'roc-raw_results.pdf')

11.2.3.24 results_analysed() `def cando.CANDO.results_analysed (`
 self,
 f,
 metrics,
 effect_type)

Creates the results analysed named file for the benchmarking and computes final avg indication accuracies.

Parameters

<i>f</i>	str: File path for results analysed named
<i>metrics</i>	list: Cutoffs used for the benchmarking protocol
<i>effect_type</i>	str: Defines the effect as either an Indication (disease) or ADR (adverse reaction)

11.2.3.25 save_rmsds_to_file() `def cando.CANDO.save_rmsds_to_file (`
 self,
 f)

Write calculated distances of all compounds to all compounds to file.

Parameters

<i>f</i>	File name to save distances
----------	-----------------------------

11.2.3.26 search_compound() `def cando.CANDO.search_compound (`
 self,
 name,
 n = 5)

Print closest [Compound](#) names/IDs for input search str.

Parameters

<i>name</i>	str: Compound name
<i>n</i>	int: Number of outputted compounds

Returns

Returns None

```
11.2.3.27 search_indication() def cando.CANDO.search_indication (
    self,
    name,
    n = 5 )
```

Print closest MeSH IDs for [Indication](#) name.

Parameters

<i>name</i>	str: Indication name
<i>n</i>	int: Number of outputted indications

Returns

Returns None

```
11.2.3.28 sigs() def cando.CANDO.sigs (
    self,
    rm )
```

Return a list of all signatures, rm is a list of compound ids you do not want in the list.

Parameters

<i>rm</i>	list: List of compound ids to remove from list of signatures
-----------	--

Returns

list: List of all signatures

```
11.2.3.29 similar_compounds() def cando.CANDO.similar_compounds (
    self,
```

```
    compd,  
    n = 10 )
```

Computes and prints the top n most similar compounds to an input [Compound](#) object `cando_comp` or input novel signature `new_sig`.

Parameters

<i>compd</i>	Compound : Compound object
<i>n</i>	int: top number of similar Compounds to be used for prediction

```
11.2.3.30 top_targets() def cando.CANDO.top_targets (  
    self,  
    compd,  
    n = 10,  
    negative = False )
```

Get the top scoring protein targets for a given compound.

Parameters

<i>compd</i>	Compound or int: Compound object or int id_ for which to print targets
<i>n</i>	int: number of top targets to print/return
<i>negative</i>	int: if the interaction scores are negative (stronger) energies

Returns

Returns list: list of tuples (protein id_, score)

```
11.2.3.31 uniprot_set_index() def cando.CANDO.uniprot_set_index (  
    self,  
    prots )
```

Gather proteins from input matrix that map to UniProt IDs from 'protein_set=' param.

Parameters

<i>perts</i>	list: UniProt IDs (str)
--------------	-------------------------

Returns

Returns list: [Protein](#) chains (str) matching input UniProt IDs

```

11.2.3.32 virtual_screen() def cando.CANDO.virtual_screen (
    self,
    protein,
    n = 10,
    negative = False,
    compound_set = 'all',
    save_file = '' )

```

Get the top scoring protein targets for a given compound.

Parameters

<i>protein</i>	Protein int or str: Protein (object, int index, or str id_) of which to screen for top scores
<i>n</i>	int: number of top compounds to print/return
<i>negative</i>	int: if the interaction scores are negative (stronger) energies
<i>compound_set</i>	str: use all Compounds ('all') or only approved Compounds ('approved')
<i>save_file</i>	str: save results to file name

Returns

Returns list: list of tuples (compound id_, score)

11.2.4 Member Data Documentation

11.2.4.1 accuracies `cando.CANDO.accuracies`

11.2.4.2 adr_map `cando.CANDO.adr_map`

str: File path to [ADR](#) mapping file

11.2.4.3 adrs `cando.CANDO.adrs`

11.2.4.4 c_map `cando.CANDO.c_map`

str: File path to the compound mapping file (relative or absolute)

11.2.4.5 clusters `cando.CANDO.clusters`

11.2.4.6 compd_set `cando.CANDO.compound_set`

11.2.4.7 compounds `cando.CANDO.compounds`

11.2.4.8 compute_distance `cando.CANDO.compute_distance`

bool: Calculate the distance for each [Compound](#) against all other Compounds using chosen distance metric

11.2.4.9 data_name `cando.CANDO.data_name`

11.2.4.10 dist_metric `cando.CANDO.dist_metric`

str: Distance metric to be used for computing Compound-Compound distances

11.2.4.11 i_map `cando.CANDO.i_map`

str: File path to the indication mapping file (relative or absolute)

11.2.4.12 indication_ids `cando.CANDO.indication_ids`

11.2.4.13 indication_pathways `cando.CANDO.indication_pathways`

str: File path to Indication-Pathway association file

11.2.4.14 indication_proteins `cando.CANDO.indication_proteins`

str: File path to Indication-Protein association file

11.2.4.15 indications `cando.CANDO.indications`

11.2.4.16 matrix `cando.CANDO.matrix`

str: File path to the cando matrix file (relative or absolute)

11.2.4.17 ncpus `cando.CANDO.ncpus`

int: Number of CPUs used for parallelization

11.2.4.18 pathway_quantifier `cando.CANDO.pathway_quantifier`

str: Method used to quantify a all Pathways

11.2.4.19 pathways `cando.CANDO.pathways`

str: File path to pathway file

11.2.4.20 protein_id_to_index `cando.CANDO.protein_id_to_index`

11.2.4.21 protein_map `cando.CANDO.protein_map`

str: File path to [Protein](#) metadata mapping file

11.2.4.22 protein_set `cando.CANDO.protein_set`

str: File path to protein subset file (relative or absolute)

11.2.4.23 proteins `cando.CANDO.proteins`

11.2.4.24 read_rmsds `cando.CANDO.read_rmsds`

str: File path to pre-computed distance matrix

11.2.4.25 rm_cmpds `cando.CANDO.rm_cmpds`

11.2.4.26 rm_compounds `cando.CANDO.rm_compounds`

list: Compounds to remove from the [CANDO](#) object

11.2.4.27 rm_zeros `cando.CANDO.rm_zeros`

bool: Remove Compounds with all-zero signatures from [CANDO](#) object

11.2.4.28 save_rmsds `cando.CANDO.save_rmsds`

bool: Write the calculated distances to file after computation (set `compute_distances=True`)

11.2.4.29 short_matrix_path `cando.CANDO.short_matrix_path`**11.2.4.30 short_protein_set** `cando.CANDO.short_protein_set`**11.2.4.31 short_read_rmsds** `cando.CANDO.short_read_rmsds`**11.2.4.32 similarity** `cando.CANDO.similarity`

bool: Use similarity instead of distance

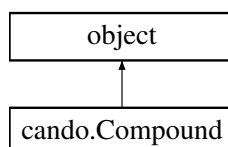
The documentation for this class was generated from the following file:

- [cando.py](#)

11.3 cando.Compound Class Reference

An object to represent a compound/drug.

Inheritance diagram for `cando.Compound`:



Public Member Functions

- `def __init__` (self, [name](#), [id_](#), [index](#), [status](#)='N/A')
- `def add_indication` (self, [ind](#))

Add an [Indication](#) to the list of [Indications](#) associated to this [Compound](#).

Public Attributes

- [name](#)
- [id_](#)
- [index](#)
- [status](#)
- [sig](#)
- [aux_sig](#)
- [indications](#)
- [similar](#)
- [similar_computed](#)
- [similar_sorted](#)
- [cluster_id](#)
- [adrs](#)
- [alt_ids](#)

11.3.1 Detailed Description

An object to represent a compound/drug.

11.3.2 Constructor & Destructor Documentation

11.3.2.1 `__init__()` `def cando.Compound.__init__ (`
 `self,`
 `name,`
 `id_,`
 `index,`
 `status = 'N/A')`

11.3.3 Member Function Documentation

11.3.3.1 `add_indication()` `def cando.Compound.add_indication (`
 `self,`
 `ind)`

Add an [Indication](#) to the list of Indications associated to this [Compound](#).

Parameters

<i>ind</i>	object: Indication object to add
------------	--

11.3.4 Member Data Documentation

11.3.4.1 **adrs** `cando.Compound.adrs`

list: List of ADRs associated with this [Compound](#)

11.3.4.2 **alt_ids** `cando.Compound.alt_ids`

dict: dict of other ids inputted with compound mapping

11.3.4.3 **aux_sig** `cando.Compound.aux_sig`

list: Potentially temporary signature for things like pathways, where "c.sig" needs to be preserved

11.3.4.4 **cluster_id** `cando.Compound.cluster_id`

int: The cluster id this [Compound](#) was assigned from clustering method

11.3.4.5 **id_** `cando.Compound.id_`

int: [CANDO](#) id from mapping file (e.g., 1, 10, 100, ...)

11.3.4.6 **index** `cando.Compound.index`

int: The order in which the [Compound](#) appears in the mapping file (e.g, 1, 2, 3, ...)

11.3.4.7 **indications** `cando.Compound.indications`

list: This is every indication the [Compound](#) is associated with from the mapping file

11.3.4.8 **name** `cando.Compound.name`

str: Name of the [Compound](#) (e.g., 'caffeine')

11.3.4.9 **sig** `cando.Compound.sig`

list: Signature is essentially a column of the [Matrix](#)

11.3.4.10 **similar** `cando.Compound.similar`

list: This is the ranked list of compounds with the most similar interaction signatures

11.3.4.11 similar_computed `cando.Compound.similar_computed`

bool: Have the distances of all Compounds to the given [Compound](#) been computed?

11.3.4.12 similar_sorted `cando.Compound.similar_sorted`

bool: Have the most similar Compounds to the given [Compound](#) been sorted?

11.3.4.13 status `cando.Compound.status`

str: The clinical trial status of the compound from DrugBank ('approved' or 'other')

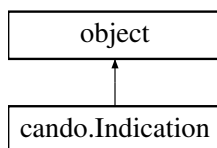
The documentation for this class was generated from the following file:

- [cando.py](#)

11.4 cando.Indication Class Reference

An object to represent an indication (disease)

Inheritance diagram for `cando.Indication`:



Public Member Functions

- `def __init__(self, ind_id, name)`

Public Attributes

- [id_](#)
- [name](#)
- [compounds](#)
- [pathways](#)
- [proteins](#)
- [pathogen](#)

11.4.1 Detailed Description

An object to represent an indication (disease)

11.4.2 Constructor & Destructor Documentation

11.4.2.1 `__init__()` `def cando.Indication.__init__ (`
 `self,`
 `ind_id,`
 `name)`

11.4.3 Member Data Documentation

11.4.3.1 **compounds** `cando.Indication.compounds`

list: Every associated compound object from the mapping file

11.4.3.2 **id_** `cando.Indication.id_`

str: MeSH or OMIM ID for the indication from the mapping file

11.4.3.3 **name** `cando.Indication.name`

str: Name for the indication from the mapping file

11.4.3.4 **pathogen** `cando.Indication.pathogen`

bool: Whether or not this indication is caused by a pathogen

11.4.3.5 **pathways** `cando.Indication.pathways`

list: Every pathway associated to the indication from the mapping file

11.4.3.6 **proteins** `cando.Indication.proteins`

list: Every protein associated to the indication from the mapping file

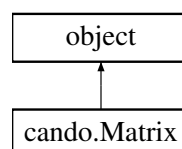
The documentation for this class was generated from the following file:

- [cando.py](#)

11.5 cando.Matrix Class Reference

An object to represent a matrix.

Inheritance diagram for `cando.Matrix`:



Public Member Functions

- `def __init__ (self, matrix_file, rmsd=False, convert_to_tsv=False)`
- `def convert (self, out_file)`
- Convert similarity matrix to distance matrix or vice versa.*
- `def normalize (self, outfile, dimension='drugs', method='avg')`
- Normalize the interaction scores across drugs (default) or proteins (not implemented yet).*

Public Attributes

- [matrix_file](#)
- [rmsd](#)
- [convert_to_tsv](#)
- [proteins](#)
- [values](#)

11.5.1 Detailed Description

An object to represent a matrix.

Intended for easier handling of matrices. Convert between fpt and tsv, as well as distance to similarity (and vice versa)

11.5.2 Constructor & Destructor Documentation

11.5.2.1 `__init__()` `def cando.Matrix.__init__ (`
 `self,`
 `matrix_file,`
 `rmsd = False,`
 `convert_to_tsv = False)`

11.5.3 Member Function Documentation

11.5.3.1 `convert()` `def cando.Matrix.convert (`
 `self,`
 `out_file)`

Convert similarity matrix to distance matrix or vice versa.

The first value in the matrix will determine the type of conversion (0.0 means distance to similarity, 1.0 means similarity to distance).

Parameters

<i>out_file</i>	str: File path to which write the converted matrix.
-----------------	---

11.5.3.2 normalize() `def cando.Matrix.normalize (`
 self,
 outfile,
 dimension = 'drugs',
 method = 'avg')

Normalize the interaction scores across drugs (default) or proteins (not implemented yet).

Parameters

<i>outfile</i>	str: File path to which is written the converted matrix.
<i>dimension</i>	str: which vector to normalize - either 'drugs' to normalize all scores within the proteomic vector or 'proteins' to normalize for a protein against all drug scores.
<i>method</i>	str: normalize by the average or max within the vectors

11.5.4 Member Data Documentation

11.5.4.1 convert_to_tsv `cando.Matrix.convert_to_tsv`

bool: Convert old matrix format (.fpt) to .tsv

11.5.4.2 matrix_file `cando.Matrix.matrix_file`

str: Path to file with interaction scores

11.5.4.3 proteins `cando.Matrix.proteins`

list: Proteins in the [Matrix](#)

11.5.4.4 rmsd `cando.Matrix.rmsd`

bool: if the matrix_file is an rmsd file

11.5.4.5 values `cando.Matrix.values`

list: Values in the [Matrix](#)

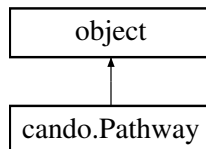
The documentation for this class was generated from the following file:

- [cando.py](#)

11.6 cando.Pathway Class Reference

An object to represent a pathway.

Inheritance diagram for cando.Pathway:



Public Member Functions

- `def __init__(self, id_)`

Public Attributes

- `proteins`
- `id_`
- `indications`

11.6.1 Detailed Description

An object to represent a pathway.

11.6.2 Constructor & Destructor Documentation

11.6.2.1 `__init__()` `def cando.Pathway.__init__ (`
 `self,`
 `id_)`

11.6.3 Member Data Documentation

11.6.3.1 `id_ cando.Pathway.id_`

str: Identification for the given [Pathway](#)

11.6.3.2 `indications cando.Pathway.indications`

list: [Indication](#) objects associated with the given [Pathway](#)

11.6.3.3 proteins `cando.Pathway.proteins`

list: [Protein](#) objects associated with the given [Pathway](#)

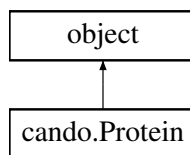
The documentation for this class was generated from the following file:

- [cando.py](#)

11.7 cando.Protein Class Reference

An object to represent a protein.

Inheritance diagram for `cando.Protein`:



Public Member Functions

- `def __init__(self, id_, sig)`

Public Attributes

- [id_](#)
- [alt_id](#)
- [sig](#)
- [pathways](#)
- [name](#)
- [gene](#)

11.7.1 Detailed Description

An object to represent a protein.

11.7.2 Constructor & Destructor Documentation

11.7.2.1 `__init__()` `def cando.Protein.__init__(
 self,
 id_,
 sig)`

11.7.3 Member Data Documentation

11.7.3.1 **alt_id** `cando.Protein.alt_id`

Used when a second identifier mapping is available (such as SIFTs project)

11.7.3.2 **gene** `cando.Protein.gene`

11.7.3.3 **id_** `cando.Protein.id_`

PDB or UniProt ID for the given protein

11.7.3.4 **name** `cando.Protein.name`

11.7.3.5 **pathways** `cando.Protein.pathways`

List of [Pathway](#) objects in which the given protein is involved.

11.7.3.6 **sig** `cando.Protein.sig`

List of scores representing each drug interaction with the given protein

The documentation for this class was generated from the following file:

- [cando.py](#)

12 File Documentation

12.1 AUTHORS.md File Reference

12.2 cando.py File Reference

Classes

- class [cando.Protein](#)
An object to represent a protein.
- class [cando.Compound](#)
An object to represent a compound/drug.
- class [cando.Indication](#)
An object to represent an indication (disease)
- class [cando.Pathway](#)
An object to represent a pathway.
- class [cando.ADR](#)
An object to represent an adverse reaction.
- class [cando.CANDO](#)
An object to represent all aspects of [CANDO](#) (compounds, indications, matrix, etc.)
- class [cando.Matrix](#)
An object to represent a matrix.

Namespaces

- [cando](#)

Functions

- def [cando.generate_matrix](#) (cmpd_scores, prot_scores, c_cutoff=0.0, p_cutoff=0.0, percentile_cutoff=None, interaction_score='P', matrix_file='cando_interaction_matrix.tsv', ncpus=1)
Generate a [CANDO Matrix](#).
- def [cando.generate_scores](#) (fp="rd_ecfp4", cmpd_pdb="", out_path='.')
Generate the fingerprint for a new compound and calculate the Tanimoto similarities against all binding site ligands.
- def [cando.generate_signature](#) (cmpd_scores="", prot_scores="", c_cutoff=0.0, p_cutoff=0.0, percentile_cutoff=None, interaction_score='P', matrix_file="")
Generate signature.
- def [cando.get_scores](#) (c, p_scores, c_score, c_cutoff, p_cutoff, percentile_cutoff, i_score)
Get best score for each Compound-Protein interaction.
- def [cando.score_fp](#) (fp, cmpd_file, cmpd_id, bs)
Generate the scores for a given [Compound](#) against all [Protein](#) ligands.
- def [cando.cosine_dist](#) (A)
- def [cando.tanimoto_sparse](#) (str1, str2)
Calculate the tanimoto coefficient for a pair of sparse vectors.
- def [cando.tanimoto_dense](#) (list1, list2)
Calculate the tanimoto coefficient for a pair of dense vectors.
- def [cando.get_fp_lig](#) (fp)
Download precompiled binding site ligand fingerprints using the given fingerprint method.
- def [cando.get_v2](#) (matrix='nrpdb')
Download [CANDO](#) v2.0 data.
- def [cando.get_tutorial](#) ()
Download data for tutorial.
- def [cando.get_test](#) ()
Download data for test script.
- def [cando.dl_dir](#) (url, out, l)
Function to recursively download a directory.
- def [cando.dl_file](#) (url, out_file)
Function to download a file.

12.3 LICENSE.md File Reference

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