An integrated family of amino acid sequence analysis programs

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Abstract

During the last years abundant sequence data has become available due to the rapid progress in protein and DNA sequencing techniques. The exact three-dimensional structures, however, are available only for a fraction of proteins with known sequences. For many purposes the primary amino acid sequence of a protein can be directly used to predict important structural parameters. However, mathematical presentation of the calculated values often makes interpretation difficult, especially if many proteins must be analysed and compared. Here we introduce a broad-based, user-defined analysis of amino acid sequence information. The program package is based on published algorithms and is designed to access standard protein data bases, calculate hydropathy, surface probability and flexibility values and perform secondary structure predictions. The data output is in an 'easy-to-read' graphic format and several parameters can be superimposed within a single plot in order to simplify data interpretations. Additionally, this package includes a novel algorithm for the prediction of potential antigenic sites. Thus the software package presented here offers a powerful means of analysing an amino acid sequence for the purpose of structure/function studies as well as antigenic site analyses. These algorithms were written to function in context with the UWGCG (University of Wisconsin Genetics Computer Group) program collection, and are now distributed within that package.

Introduction

Nucleic acid and protein sequences have been evaluated and are available for most laboratories by accessing sequence libraries. Precise information about the structures of these proteins can be reliably obtained by methods such as X-ray diffraction (Liljas and Rossman, 1974) and two-dimensional NMR spectroscopy (Wagner and Wüthrich, 1982); these methods, however, require relatively large amounts of pure crystalline polypeptides and can be applied only to a very small number of proteins with known primary amino acid sequences. A great deal of information, however, may also be obtained for many Downloaded from bioinformatics.oxfordjournals.org at Universitaetsbibliothek Regensburg on April 14, 2011

proteins by a careful examination of the linear amino acid sequence. For these reasons computer algorithms were developed for analysis of such sequences, enabling the prediction of probable secondary structural features (Chou and Fasman, 1978; Garnier *et al.*, 1978), regional backbone flexibility (Karplus and Schulz, 1985) as well as parameters relating to surface accessibility (Janin *et al.*, 1978). The subsequent application of these predictions may facilitate the identification of functional and structural parameters, i.e. transmembranespanning regions and signal peptides and may lead to a better understanding of the three-dimensional arrangement of a given amino acid sequence.

Here we present an integrated protein analysis software package which uses the algorithms of Chou and Fasman (1978) and Garnier et al. (1978) for prediction of the secondary structures. Since numerical presentation of the calculated values is often rather difficult to interpret, the data may be presented in a two-dimensional or linear graphic output. These structural values may be superimposed with additional parameters such as hydrophilicity (Hopp and Woods, 1981; Kyte and Doolittle, 1983), flexibility (Karplus and Schulz, 1985) and surface probability (modified from Emini et al., 1985). The combination of these parameters with the secondary structure predictions facilitates the identification of continuous antigenic sites in an amino acid sequence. Such structures are most often located in regions with a high content of β -turns and/or high values for hydrophilicity, flexibility and surface probability; the immunodominant antigenic determinants generally exist in looplike structures at the surface of a protein molecule. As a first step towards automated prediction of antigenic sites, these parameters were combined in a weighted manner resulting in a novel algorithm, the antigenic index (Jameson and Wolf, 1988).

Systems and methods

The programs PROTCALC for calculation of secondary protein structures and additional parameters such as hydrophilicity, surface probability, flexibility, antigenicity and potential Nglycosylation, and PROTPLOT for graphic design were written in VAX/FORTRAN (version 4.1); the VAX 750 (Digital Equipment Corporation) was used as host computer. All drawings were produced on a Hewlett Package Plotter 7475A. The protein sequences were taken from the GENBANK sequence data library, EMBO sequence collection or NBRF protein data

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bank. The algorithms were written to function within the UWGCG Sequence Analysis Software Package (Devereux *et al.*, 1984); both the analysis program and the graphics output are part of this program collection.

Algorithm

Algorithms for the prediction of secondary protein structure were taken from Garnier *et al.* (1978) and Chou and Fasman (1978). The overlapping regions of an α -helix and β -sheet were resolved by using the 'overall probability' introduced by Nishikawa (1983). The same procedure was also applied to locate turn regions which are inconsistent with other secondary structures. The following modifications of Chou – Fasman rules were used for α -helical regions: the boundary conditions of p(bound) > 1.0 and necessary conditions of p(α) > p(β) are removed.

The values of hydrophilicity were determined according to Hopp and Woods (1981) or Kyte and Doolittle (1983). The latter values were multiplied by -1 to allow the same orientation of peak values as with the calculations according to Hopp and Woods and in order to facilitate the usual interpretation, i.e. that in all graphs positive or higher numeric values favor increased potential presence at the surface of a protein structure.

The calculation for the backbone flexibility of the amino acid sequence was performed as described by Karplus and Schulz (1985).

Surface probabilities were based on the individual amino acid data obtained by Janin *et al.* (1978) and calculated using a modification of the algorithm by Emini *et al.* (1985). In the equation below the surface probability at position n is defined for sequential hexapeptide sequences as

$$S_n = (\prod_{i=1}^{A} \delta_{n+4-i}) * (0.37)^{-x}$$

where δ_n is the fractional surface probability and x has the value of 6 for values of n further away from the ends than three amino acids and decreased as n approaches the ends.

The antigenic index was calculated from an experimentally derived equation based on the data derived from hydrophilicity (H), flexibility (F), surface probability (S), Chou-Fasman secondary structure prediction (CF) and Garnier secondary structure prediction (RG):

 $A_n = 0.3(H_n) + 0.15(S_n) + 0.15(F_n) + 0.2(CF_n) + 0.2(RG_n)$

(Values for H_n , S_n , F_n , CF_n and RG_n are given in Table I.) N-glycosylation sites are indicated by the sequences Asn-X-

Ser or Asn-X-Thr, and with minor probability, when X is represented by amino acids Asp, Trp or Pro.

Implementation

PROTCALC

The operator must enter the amino acid sequence which is to be analysed in the single letter code through the keyboard or—in

Values used for calculation of	Values calculated according
AI for calculated values of	to references listed
column 2	above
$H_i = 2$	H > 0.5
$H_i = 1$	0.5 > H > O
$H_i = -1$	O > H > -0.4
$H_i = -2$	-0.4 > H
$S_i = 1$	1.0 > S
$S_i = 0$	Otherwise
$\begin{aligned} F_i &= 1\\ F_i &= 0 \end{aligned}$	1.0 > F Otherwise
$CF_i = 2$	CF = strong turn
$CF_i = 1$	CF = weak turn or random coil
$CF_i = 0$	Otherwise
$RG_i = 2$	RG = strong turn
$RG_i = 1$	RG = weak turn or random coil
$RG_i = 0$	Otherwise

H, hydrophilicity; S, surface probability; F, backbone flexibility; CF, secondary structure prediction; RG, secondary structure prediction; AI, antigenic index.

the case of an already published sequence-must retrieve it from one of the sequence libraries. PROTCALC calculates the values by running a window of seven (default setting) amino acids from the beginning to the end of the protein sequence; the parameters obtained for every residue in this window are averaged; the size of the window may be altered. A further option is given in order to broaden the peaks of the antigenic index (from n - 4to n + 4) by adding 80, 60, 40 or 20% of the peak value to the flanking values in descending order to account for the influence of the additional free energy derived from the mobility of surface regions relative to regions buried inside the protein. The output of PROTCALC is presented as tables with the numerical values for hydrophilicity, surface probability, flexibility and antigenic index. Glycosylation sites of high and low probability are indicated by G and g respectively. Secondary structure parameters are given by H (α -helix), B (β -sheet) or T (β -turn) or by h, b or t for weak secondary structure parameters; random coil regions are not indicated by a special letter. The printout of the calculation for p17 of the human immunodeficiency virus 1 (HIV/HTLV-3, BH10) (Ratner et al., 1985) is shown in Figure 1.

PROTPLOT

This program uses the protein calculation file from PROTPLOT and draws a colored two-dimensional or linear graph of p17 of HIV-1 (Figure 2a and b). In the linear plot all derived parameters derived from PROTCALC may be presented in a combined graphic output. Alternatively, the operator may select single parameters to be plotted. The y axis may be enlarged by a factor of two in both cases. The operator can also choose

Table I. Computation of the antigenic index

BOTSTRUC		wh3 nhrf	check	2212. 6	rom 1	to: 132			61	τ.		+0 286	0 283	0 485	в	Ŧ	0 100
	01. 100	wii5.ii011	check.		10	10. 152			62	ž	•	-0.696	0.703	0 6 74	ě	÷	-0.200
									02		•	-0.000	0.703	1.004			-0.200
ayarophi i	ICITY (H	opp-wood	is) avera	ged over	a windo	wor: /			03	ų.	•	-0.714	1.700	1.004		1	0.100
surface P	robabili	ty accor	aing to	Emini					64	L.	•	-0.414	2.091	1.015	в	•	-0.300
Chain Fle	xibility	accordi	ing to Ka	rplus-Sc	hulz				65	Q	•	-0.414	3.031	1.021	•	•	-0.300
Secondary	Structu	re Predi	ction ac	cording	to Chou-	Fasman			66	P	•	-0.386	3.031	1.023	т	•	0.400
Secondary	Structu	re Predi	ction ac	cording	to Robso	n-Garnie	r		67	s		-0.471	4.942	1.026	т		0.100
Antigenic	ity Inde	x accord	iina to J	ameson-W	lolf				68	L		-0.214	1.762	1.033			0.000
									69	0		-0.200	1.437	1.052	_		0.002
									70	Ŧ	•	0 220	2 334	1 076	Ť		1 000
									/0	÷	•	0.614	E 767	1.006		•	1 200
Position	AAÇode	GlycoS	HyPhil	SurfPr	FlexPr	CF-Str	RG-Str	AI-Ind	/1	G	•	0.014	5.707	1.090	÷.	•	1.300
									72	5	•	0.614	2.317	1.100	n	•	0.900
1	м		0.300	1.153	ι.000			0.600	73	E	•	1.014	4.876	1.082	h	•	0.900
2	G		0.140	0.866	1.000			0.450	74	E	•	1.114	8,358	1.057	h	•	0.900
3	A		0.167	0.983	1.000	h		0.450	75	L	•	0.857	5.493	1.030	h	т	1.300
Ā	R	-	-0.071	0.696	1.000	h		-0.150	76	R		0.486	4.446	1.011	h	T	1.000
-		•	-0.143	0 794	0 066	5	•	-0.200	77	S		0.086	0.398	1.000	h	т	0.700
5	÷	•	-0.145	1 104	0.900		•	-0.300	78	L		-0.400	0.648	0.992	h	т	-0.200
9	<u> </u>	•	-0.100	1.104	0.903		•	-0.150	70	- v		-0 357	0 162	0 986		Ť	0.100
7	v	•	-0.029	0.306	0.974	n	•	-0.300	60	÷.	•	-0.337	0.107	0.900		÷	-0.200
8	L	•	+0.457	0.270	1.005	n	•	-0.450	80		•	-0.057	0.107	0.964		:	-0.200
9	s		0.043	0.438	1.037	т		0.850	81	1	•	-0.937	0.1/5	0.978		P	-0.000
10	G		-0.257	0.510	1.064	т		0.250	82	v	•	-0.957	0.088	0.967	B	в	-0.600
11	G		0.386	1.121	1.074	t		0.800	83	A	•	-0.957	0.792	0.952	в	в	-0.600
12	F		1.071	2.527	1.063	н		0.900	84	т		-1.129	0.273	0.935	B	в	-0.600
12	ĩ	•	0 543	3 579	1 041		H	0 900	85	L		-1.286	0.273	0.921	B	в	-0.600
14		•	0 971	0 064	1 029			0.000	86	Ŷ		-1.143	0.498	0.912	B	B	-0.600
14	5	•	1 400	3.904	1.028		n	0.900	87	ĉ	•	-1 043	0 760	0 021	, in the second s	ñ	-0.600
15	к	•	1.400	15.048	1.017	н	н	0.900	07	č	•	-1.043	2 607	0.921	5		-0.460
16	w	•	0.714	12.431	1.013	н	н	0.900	88	v	•	-0.557	2.007	0.938	в		-0.430
17	E	•	1.400	19.426	1.013	н	н	0.900	89	н	•	-0.557	1.077	0.961	н	н	-0.450
18	к		0.714	5.662	1.006	н	н	0.900	90	Q	•	0.200	4.738	0.984	н	н	0.450
19	I		0.714	15.451	0.995	н	н	0.750	91	R	•	0.086	4.558	0.993	н	н	0.450
20	R		1.200	11.668	0.989	н	н	0.750	92	I		0.729	9.260	0.999	н	н	0.750
21	L	-	0.771	2.775	1.000	н		0 750	93	Е		1.229	8.168	1.002	н	н	0.900
22		•	0 343	2 982	1 026		•	0.600	94	I		1.143	3.881	1.014	н	н	0.900
23	5	•	1 029	3 244	1 050		-	1 700	95	ĸ		1.143	17.533	1.034	н	н	0.900
23		•	1.029	12 105	1.039		T	1.700	06	ĥ	•	1 929	17 533	1 047	 H		0.900
24		•	1.029	12.105	1.091	T	т	1.700	07	ř	•	1 320	21 379	1 053			0.900
45	G	•	1.714	13.166	1.105	т	т	1.700	97	, i	•	1 320	5 770	1 044			0.900
20	ĸ	•	0.95/	14.111	1.096	t	т	1.500	98		·	1.329	5 720	1 029	л и		0.900
27	ĸ	•	1.386	59.326	1.081	t	т	1.500	99	-	•	1.329	5.725	1.020			0.900
28	к		1.129	66.832	1.056		т	1.300	100	•	•	1.329	13.112	1.018	н	н	0.900
29	Y		1.557	66.832	1.033		в	0.900	101	L	•	1.129	2.903	1.018	н	н	0.900
30	ĸ		1.057	32.897	1.021		в	0.900	102	D	•	1.129	2.903	1.028	н	н	0.900
31	L		0.371	7.282	0.999	в	В	0.450	103	к		1.129	7.121	1.040	н	н	0.900
32	к		-0.271	3.127	0.982	в	B	-0.150	104	I		1.629	17.595	1.049	н	н	0.900
33	н		-0.429	1.053	0.951	8	8	-0.450	105	E		1.914	19.947	1.065	н	н	0.900
34	Ť		-0 929	1 061	0 020			-0 450	106	E		1.514	1,181	1.081	н	н	0.900
35	v	•	-0 629	0 433	0 011			-0.400	107	E		1.514	5.337	1.097	н	н	0.900
36		•	-0.029	0.433	0.911			-0.600	108	õ		1.814	3.286	1.117	н	н	0.900
30	*	•	-0.629	0.808	0.914	в	н	-0.600	100	ž	÷	1 814	4 963	1 122			0 900
37	<u>^</u>	•	-0.129	2.41/	0.947	8	н	-0.150	110			1 914	7 496	1 121	÷	ü	1 300
38	S	•	-0.129	2.815	0.981	н	н	-0.150	110		•	1 814	11 239	1 110	÷	л ч	1.300
39	ĸ	•	0.514	5.532	1.004	н	н	0.900	111	3	•	1 714	51 210	1 104			0.000
40	E	•	1.429	18.846	1.019	н	н	0.900	112	ĸ	•	1.714	51.219	1.104			0.900
41	L		1.143	11.442	1.011	н	н	0.900	113	к	•	1.714	34.103	1.090	н	н	0.900
42	E		1.029	3.359	0.997	н	н	0.750	114	ĸ	•	1.314	55.880	1.073	н	н	0.900
43	R		0.386	1.167	0.977	н	н	Û.450	115	λ.		1.200	15.082	1.048	н	н	0.900
44	F		-0.014	0.258	0.957	н	н	-0.300	116	Q		0.700	4.071	1.023	н	н	0.900
45	, i		0.243	0.195	0.953	н	н	0.300	117	Ó		0.200	1.099	0.996	н	н	0.450
46	v		-0 186	0.050	0 963	н		-0 300	118	Ā		0.200	2.395	0.976	н	н	0.450
47	Ň	•	-0 971	0.055	0.000		•	-0.600	119	Ä		0.214	1.569	0.974	н	н	0.450
49		•	0.071	0.053	1 000	÷	•	0.000	120		-	0 196	0 559	0.990	и	н	0.300
40		•	-0.771	0.054	1.000	÷	•	-0.200	120	ŝ	•	0.096	1 020	1 013	ü		0.600
50		•	-0.2/1	0.156	1.007		•	0.250	121	5	•	0.000	1.020	1 0 20	÷	•	1.000
30	- L	•	-0.114	1.150	1.007	н	•	0.000	122	т	•	0.200	1.341	1.030		•	1.000
21	L	•	-0.100	0.938	1.018	н	•	-0.150	123	G	•	0.314	2,329	1.045	т	•	1.000
52	E	•	0.329	Z.610	1.040	н	•	0.600	124	н	•	0.414	2.640	1.050	•		0.600
53	т	•	0.329	2.317	1.064	н	т	1.000	125	s	•	-0.229	1,390	1.053	т		0.400
54	S		0.443	1.301	1.078	н	Т	1.000	126	S		-0.129	2.384	1.058	т		0.400
55	E	-	1.129	1.807	1.071	н	т	1.300	127	Q		-0.100	3.230	1.057	в		0.000
56	G		0.729	2.758	1.052		т	1,300	128	v		0.000	0.469	1.055	в		0.450
57	с		0.529	1.497	1.022	в	т	1.300	129	s		-0.371	0.617	1.000	в		-0 150
58	Ř		0.229	0.606	1.000	Ē	Ť	0.700	130	õ		-0.483	0.332	1.000	в	Ť	-0.050
59	ö	•	-0.200	0.606	0.987	Ř	÷	0 100	131	Ň	•	-0.620	0.518	1 000	Ř	÷	-0.050
60	Ť	•	-0.171	2.684	0 070			0.100	1 3 3		•	-0 400	0 457	1 000	5		-0.050
50	-	•	· · · · ·		0.779		1	0.230	134	1		0.400	0	1.000		L. 1	-0.050

Fig. 1. Printout of protein analysis of p17, a core protein of the human immunodeficiency virus (HIV-1).

between a one- or a four-color plot. In the two-dimensional display the secondary structure predictions by either Chou-Fasman (1978) or Garnier *et al.* (1978) are plotted; these drawings may be superimposed on the values for hydrophilicity and hydrophobicity, flexibility, surface probability or antigenic index. Those additional values are indicated by special symbols (circles or diamonds) which have been superimposed on the secondary structures (for explanation of the symbols see Figure 2a). Display of potential N-glycosylation sites may be suppressed; the default numbering is set with an interval of 50 residues but may be user-defined. There is an extra option for plotting weak Chou-Fasman parameters. Particular amino acids can be marked by their single letter code.

The two-dimensional plot is first drawn on an imaginary plane and then adjusted to give maximum filling of the given sheet format. Adjustment is done proportionally for the x and y axes.

Discussion

The two-dimensional plot allows for a fast screening of large numbers of proteins for structural parameters such as transmembrane regions or potential segments of genes likely to be recognized by the immune system. The latter feature is of particular importance when subclones, e.g. for use as an antigen source for diagnosis (Motz *et al.*, 1986), or the respective synthetic peptide are prepared and used similarly. It should be noted that the given structure in the graphic output is simply a consequence of predicted turns with all other structural elements being represented in a modified sine/zig-zag/waved line and does not resemble in any way the natural structure. However, the graphic representation, especially when superimposed values are displayed, facilitates and accelerates analysis of the gross properties of a protein.



Fig. 2. (a) Two-dimensional graph of p17 of HIV-1 using strong Chou-Fasman parameters. α : α -helical structures: ω : β -sheet; \mathfrak{D} : β -turn; α : random coil; δ : glycosylation site; \mathfrak{O} : hydrophilic regions; \diamond : hydrophobic regions. (b) Linear graph of p17 of HIV-1.

A disadvantage of this system is the variable scale which makes it difficult to compare proteins of different sizes from the sequence of structural elements where one might reflect a subunit of the other. In addition, only a limited number of parameters can be given and judged with any one drawing. These problems can be alleviated by a linear output with a fixed scale and almost unlimited space for presentation of parameters.

Those predictions and calculations have been successfully used for immunological questions, especially in the selection of peptides and protein subunits for production by chemical or gene technological methods (Motz *et al.*, 1986; Modrow and Wolf, 1986; Modrow *et al.*, 1987; Jameson *et al.*, 1987). In particular, when larger genes or genomes need to be analysed, random synthesis of peptides or shotgun expression are both expensive and time consuming. Therefore selection of specific sites is a useful approach.

Correlation with the natural conformation can only be expected to the extent which has been reported for CF and RG predictions, but in comparison with known related structures such approaches should become more reliable.

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